



ИНСТИТУТ
ДЕТСКОЙ ГЕМАТОЛОГИИ
и ТРАНСПЛАНТОЛОГИИ
имени Р. М. Горбачевой

Infections in immunocompromised patients

First State Pavlov Medical University of Saint-Petersburg, Russia; Raisa Gorbacheva Memorial Institute for Children Oncology, Hematology and Transplantation



Goloschapov O.V.

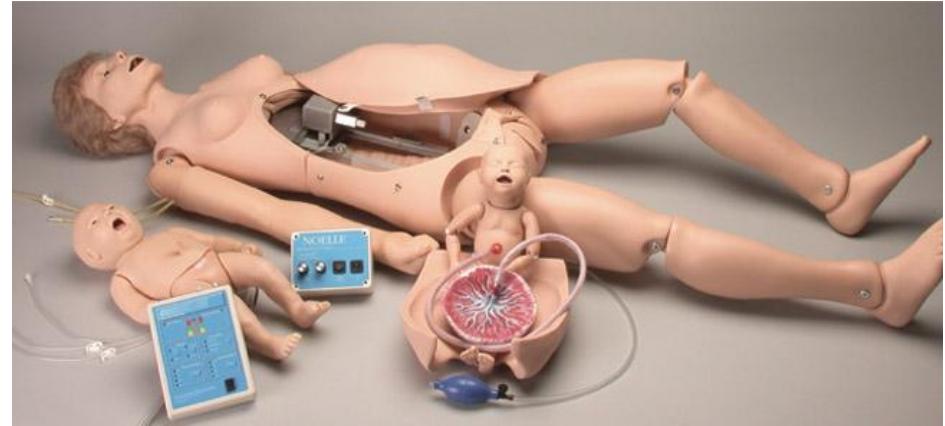
2019
САНКТ-ПЕТЕРБУРГ



Model of an immunocompromised patient?

Secondary Immunodeficiency - Immunocompromised Patient

HOW TO CREATE? How do you look? HOW TO TRAIN?



Immunocompromised patient. WHERE IS HE?

Causes of immunological failure

Allergic reactions

High microbial load

Eating disorder

- BEN
- Hypovitaminosis
- Micronutrient deficiency
- Obesity

Anatomical defects

Chronic diseases

- sakh. diabetes
- CRF
- metabolic disorders
- tumors

Iatrogenic factors

- extensive operations

Defects in the immune system

- Lekozy
- Lymphomas
- Hich

Poisoning

Burns

PREMORBID CONDITION - the state of the organism preceding the development of the disease = Condition before HSCT

Numerous infectious episodes

History of invasive mycoses (aspergillosis)
Transferred BMI, pneumonia, sinusitis

Virus infection - herpes group, polyomavirus

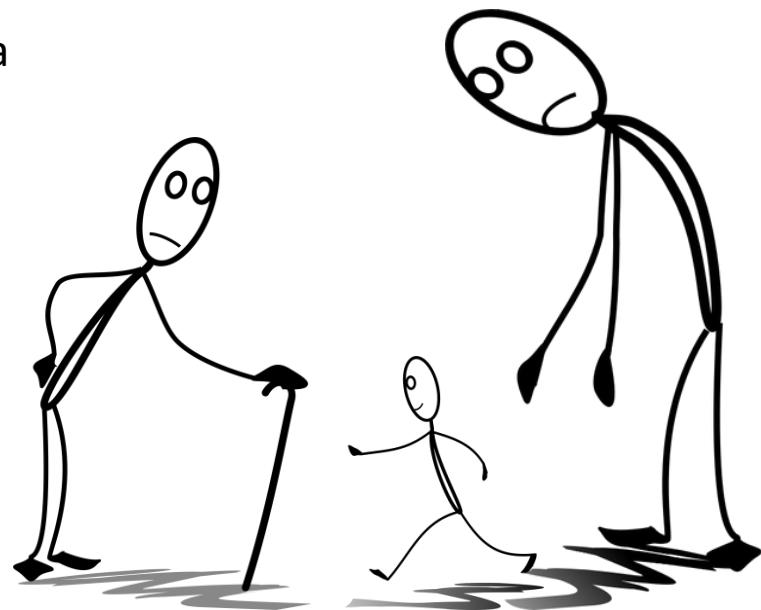
Eating disorders - low BMI, high BMI, cachexia

Genetic diseases - a violation of the metabolism of drugs, a violation of enzyme systems.

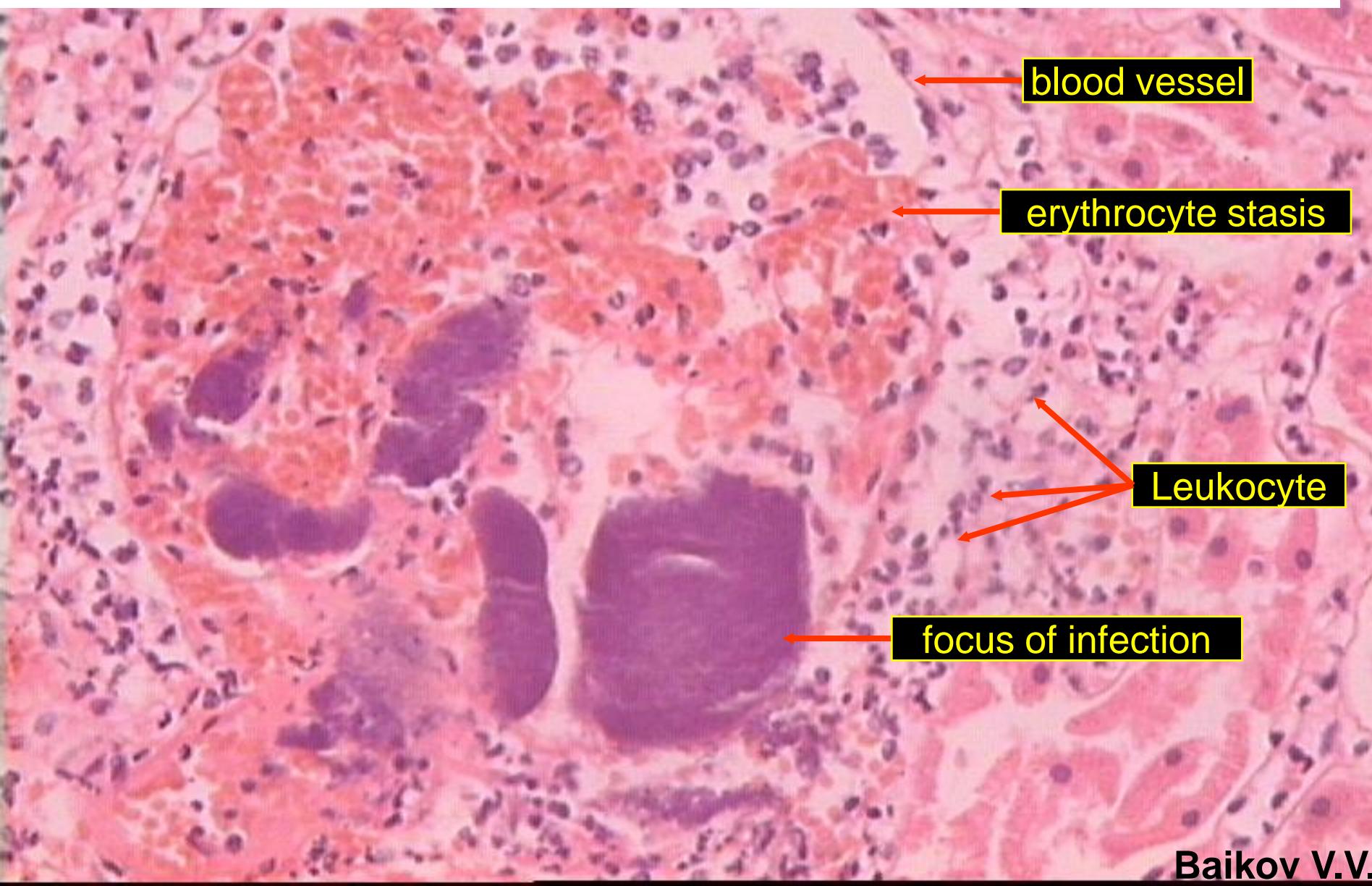
Change in PD, PK drugs.

Relapse therapy (PCT, monoclonal a / t)

Age from 0 to 90

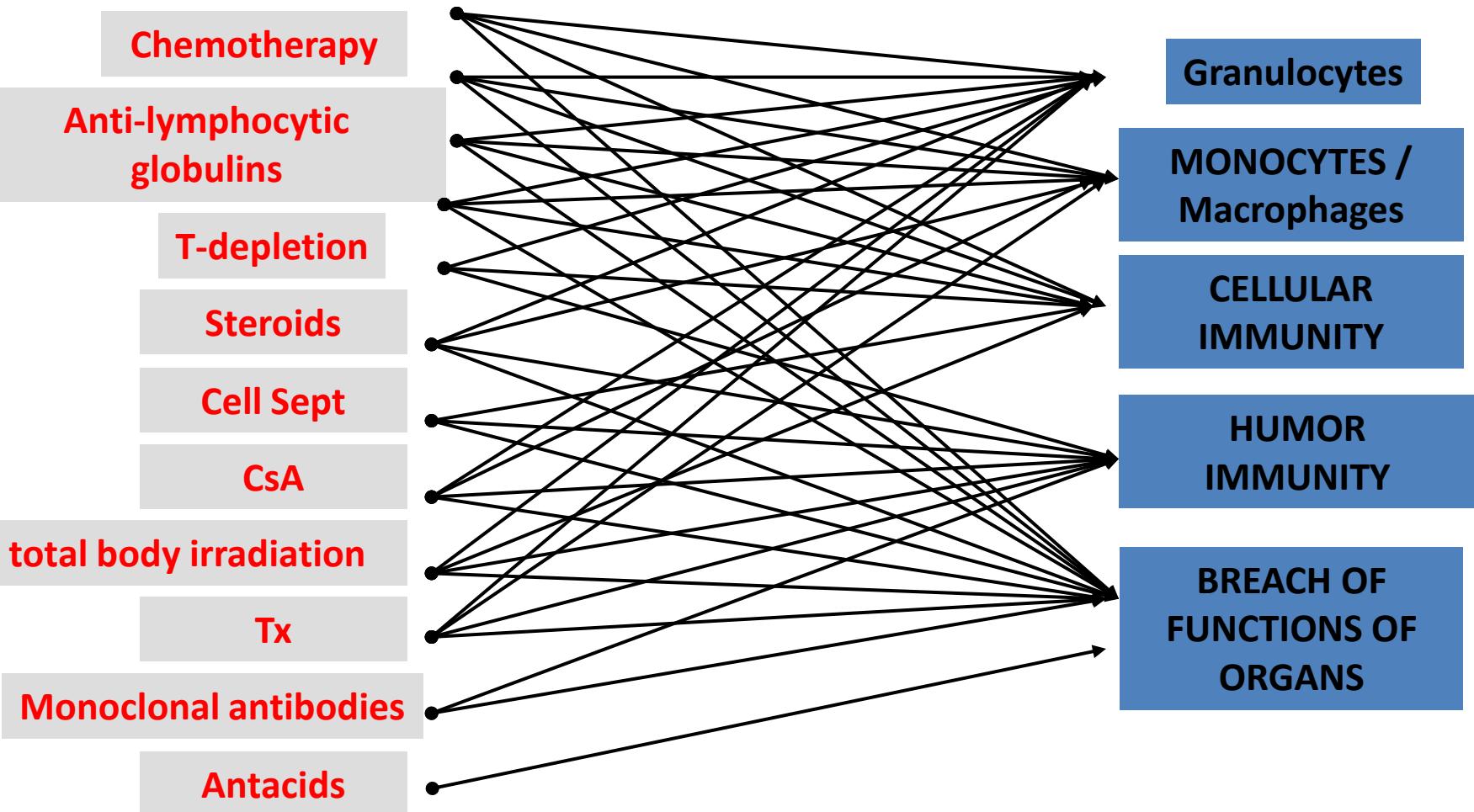


Alternative exudative path tissue damage



Risk factors for developing an oncohematological patient

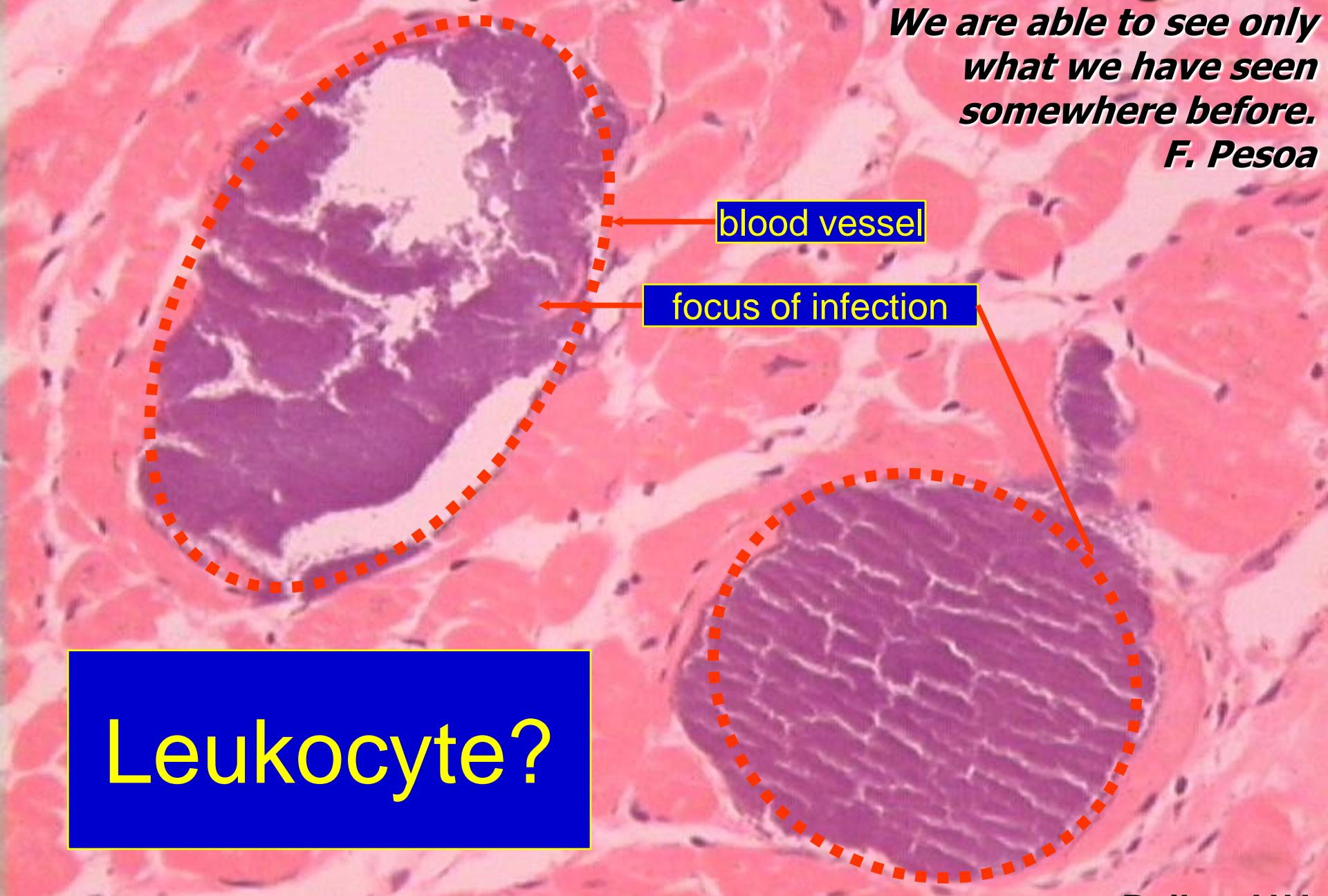
Patient model after BMT



HOW TO CREATE? - WE ALREADY CREATED (((

Alternative pathway of tissue damage

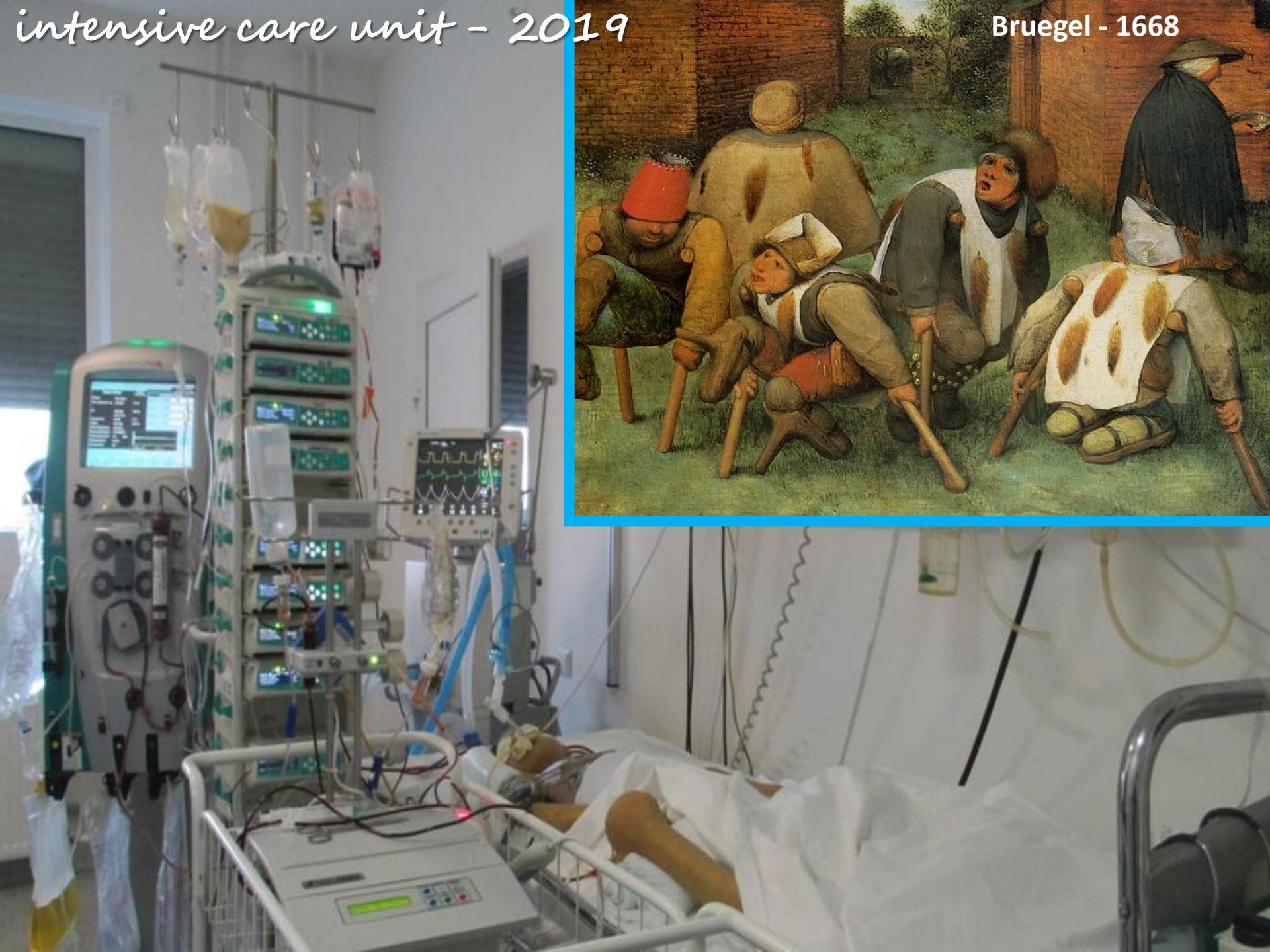
*We are able to see only
what we have seen
somewhere before.
F. Pessoa*



Leukocyte?

intensive care unit - 2019

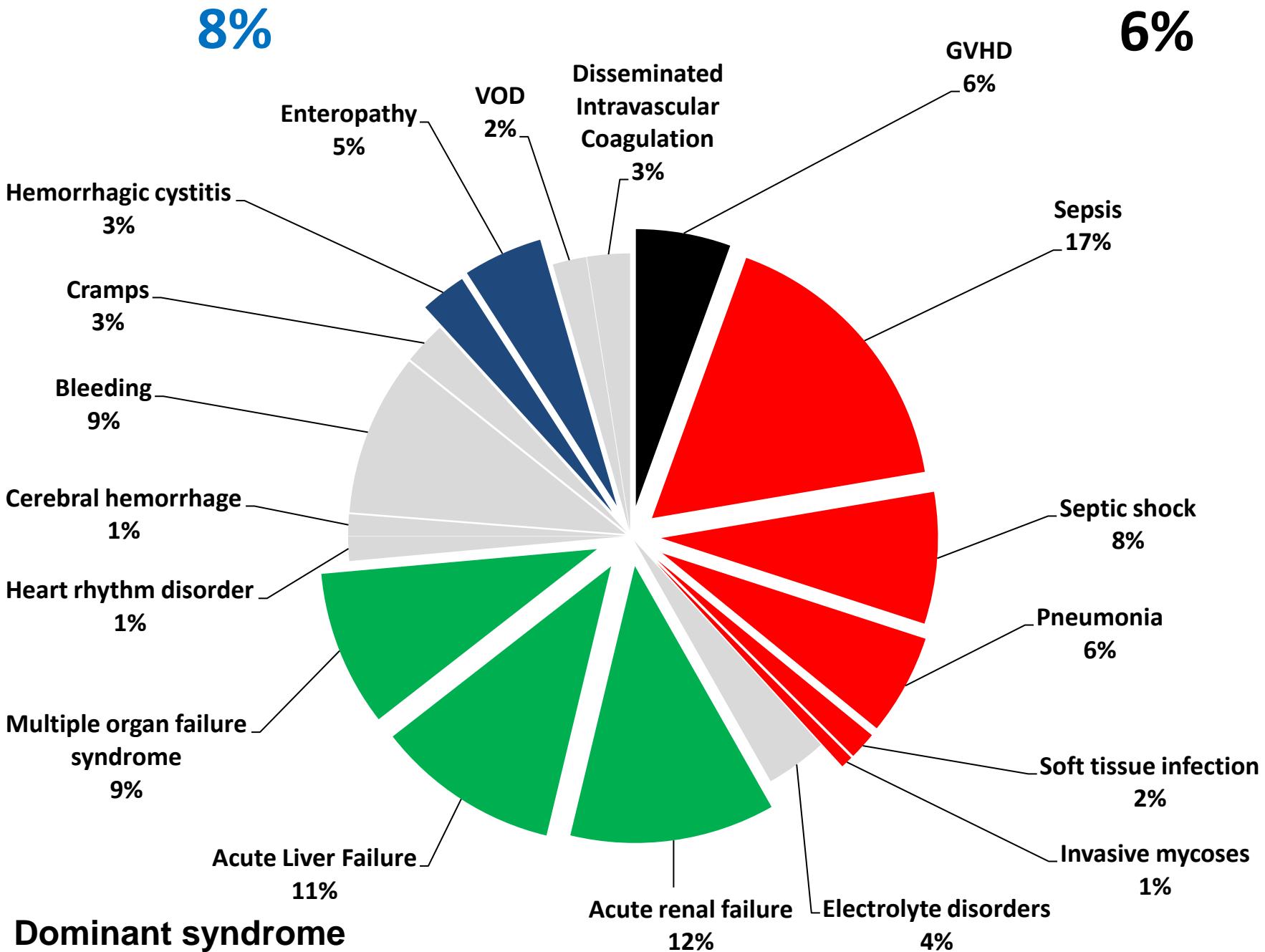
Bruegel - 1668



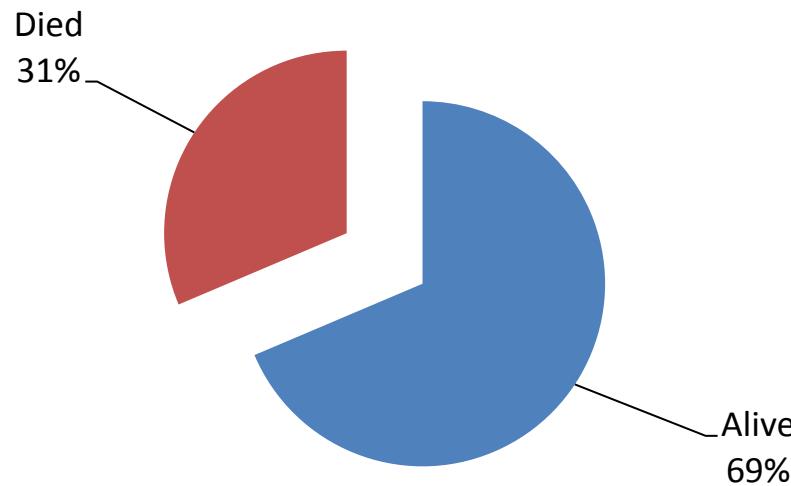
32%

Reasons for transfer to the intensive care unit 2018 (%)

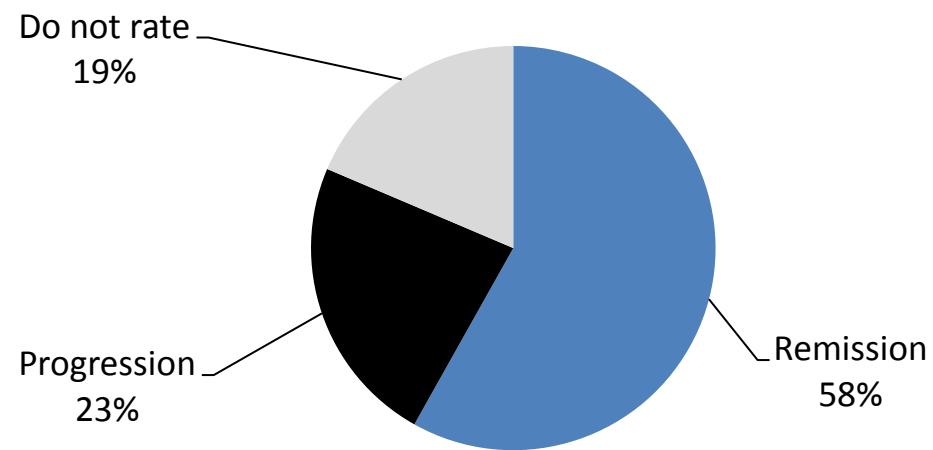
34%



Mortality of patients with sepsis 2018 (%)

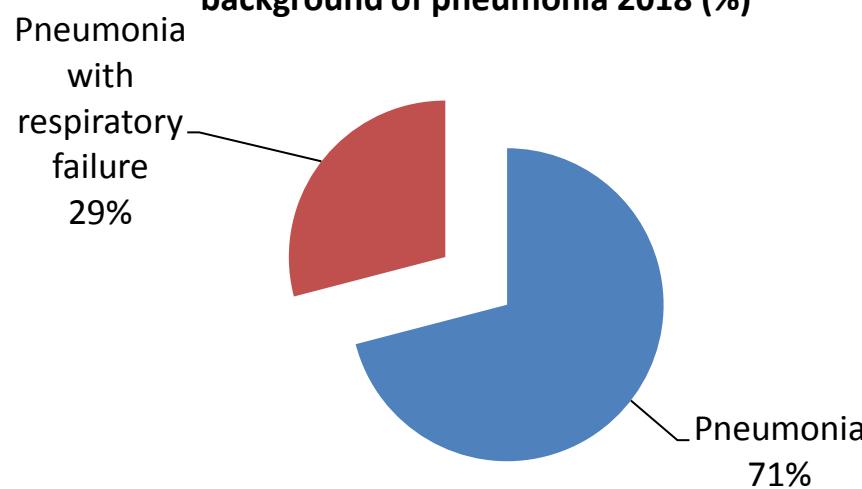


The frequency of progress of the underlying disease 2018 (%)

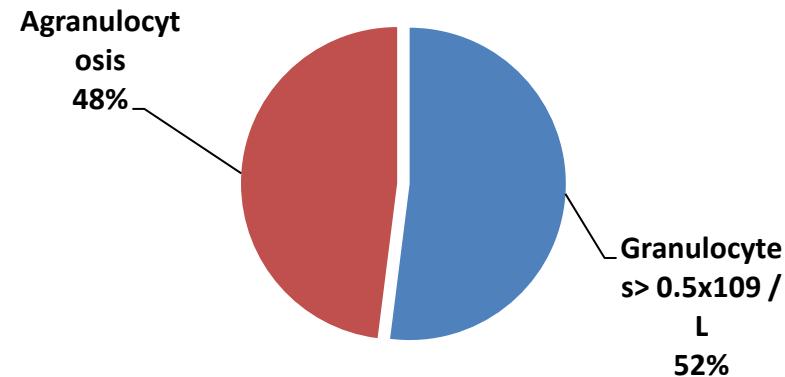


Mortality rate from sepsis after BMT 55.1% (Kumar G.et.al 2015)

The frequency of respiratory failure on the background of pneumonia 2018 (%)

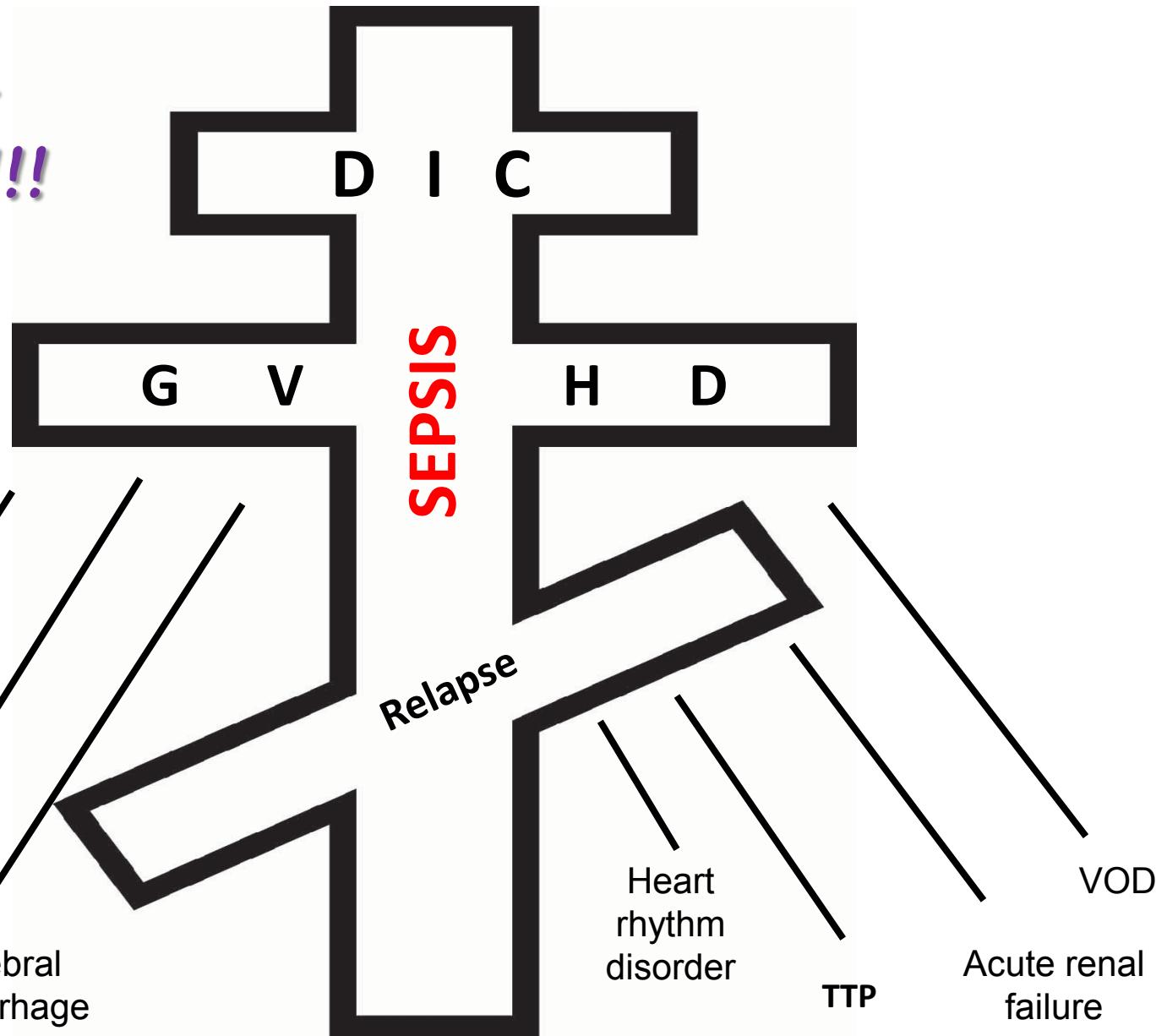


The frequency of patients with agranulocytosis 2018 (%)

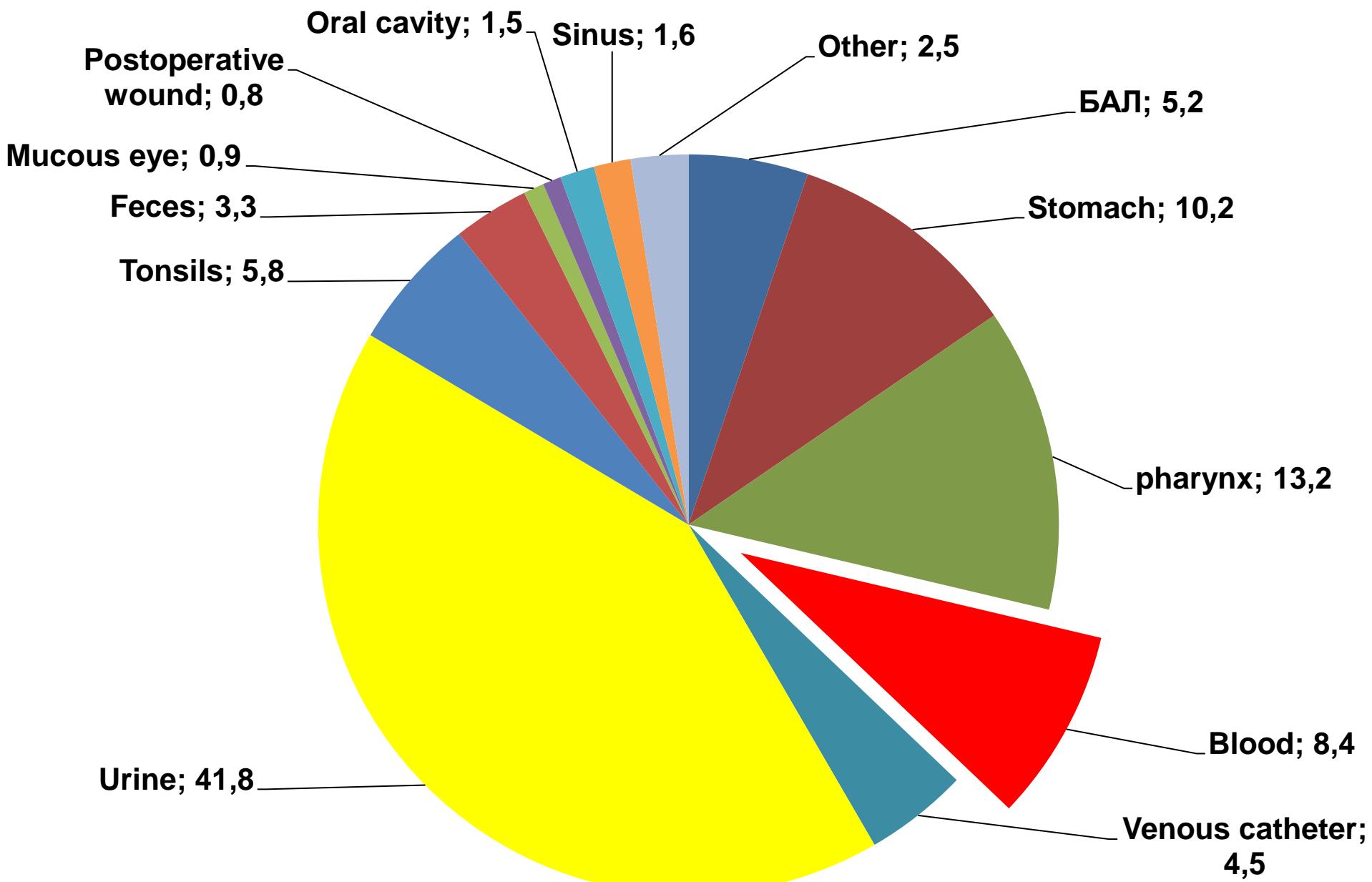


CROSS - HEMATOLOGIST

31% -
Sepsis !!!

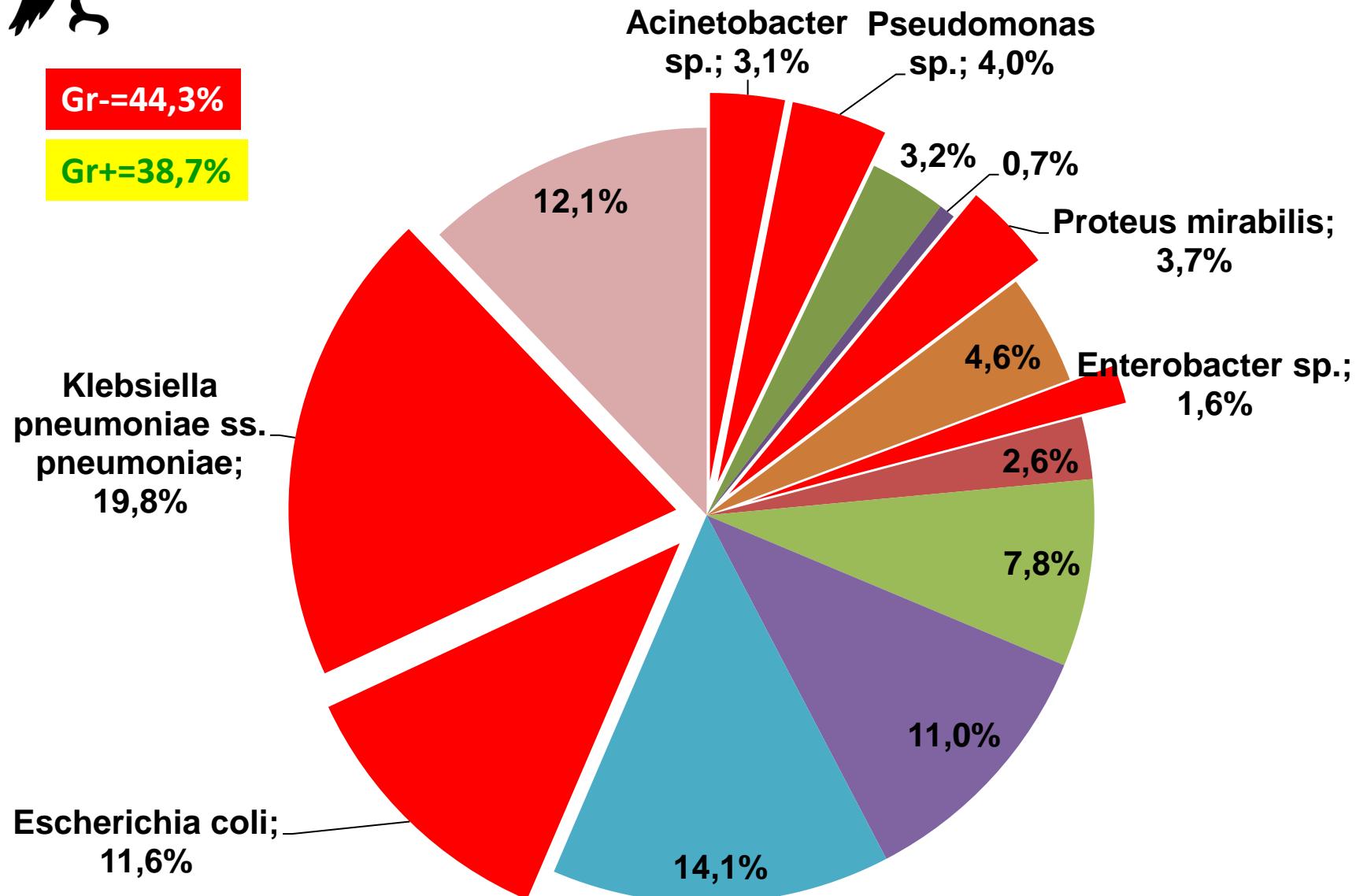


Structure of biological material from which microorganisms were isolated (%) 2018



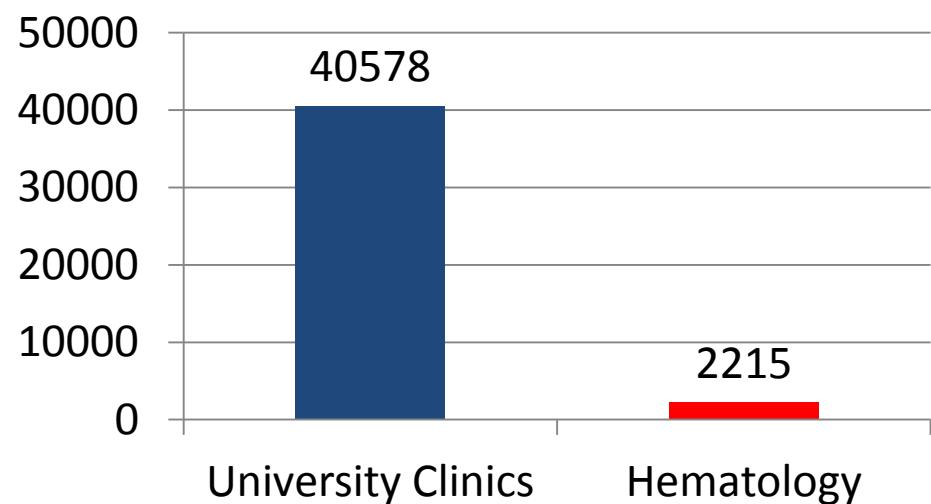


The structure of microorganisms isolated from biological material (%) 2018

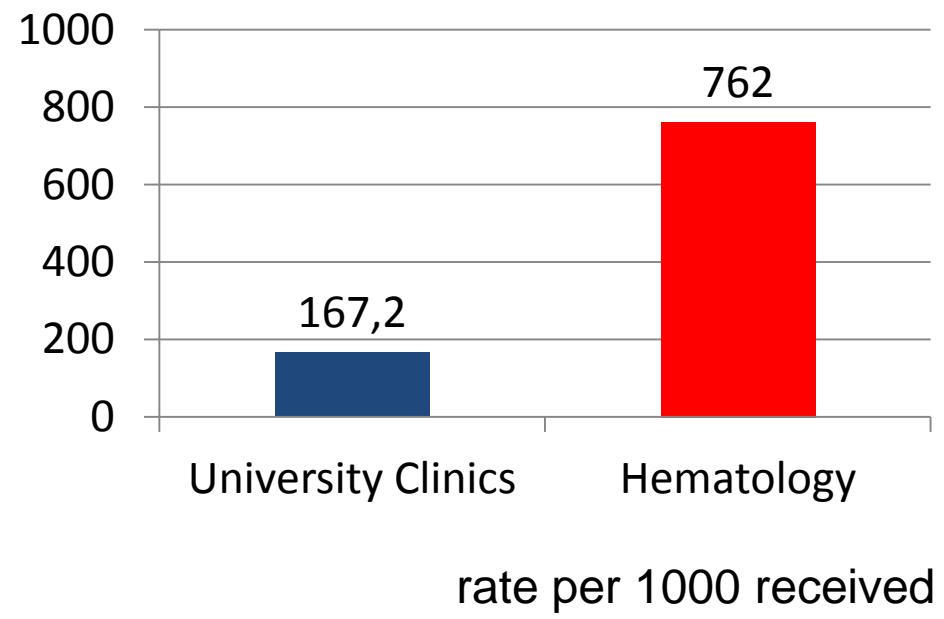


Bacteriological studies 2 times a week

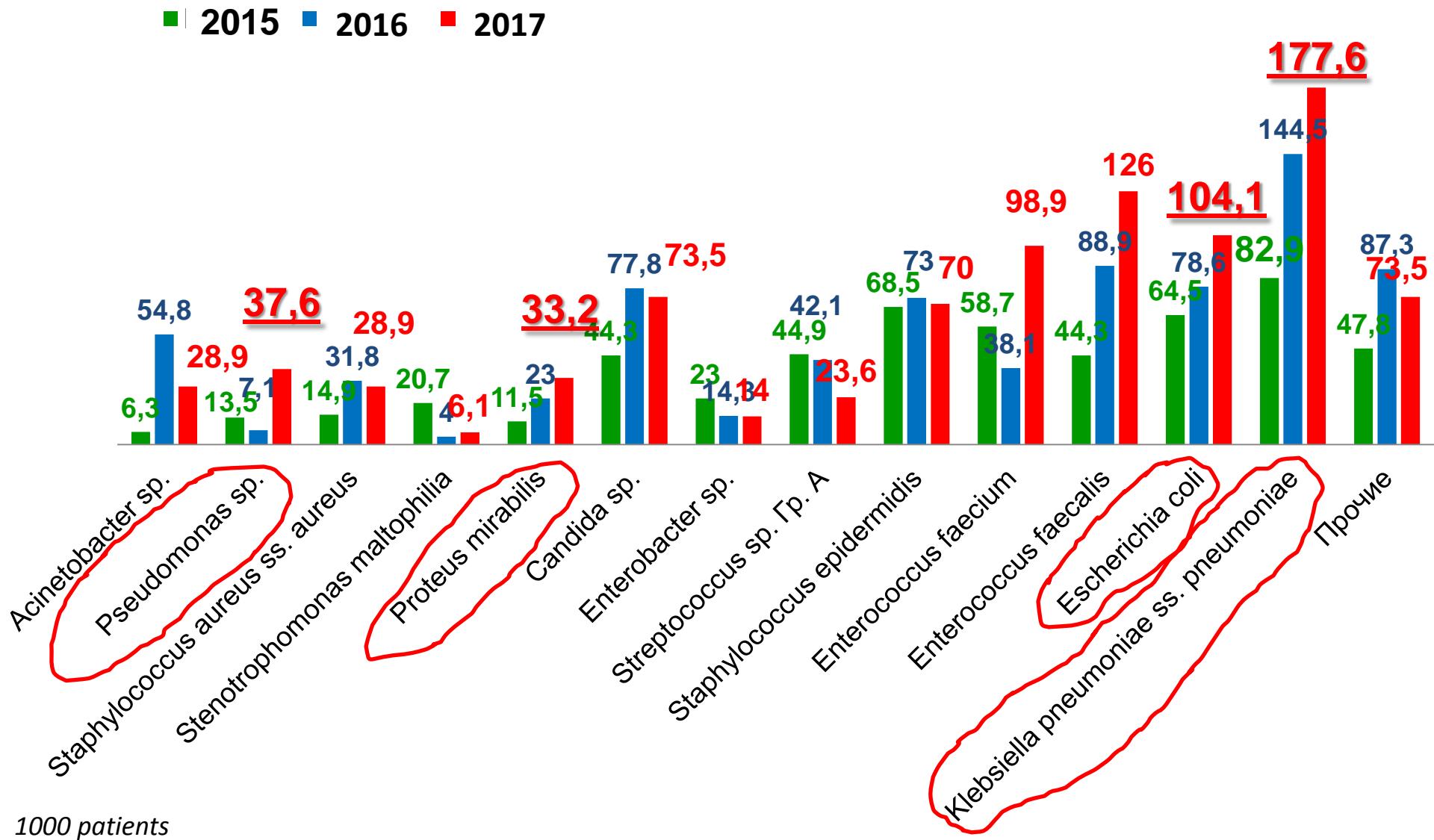
Total received



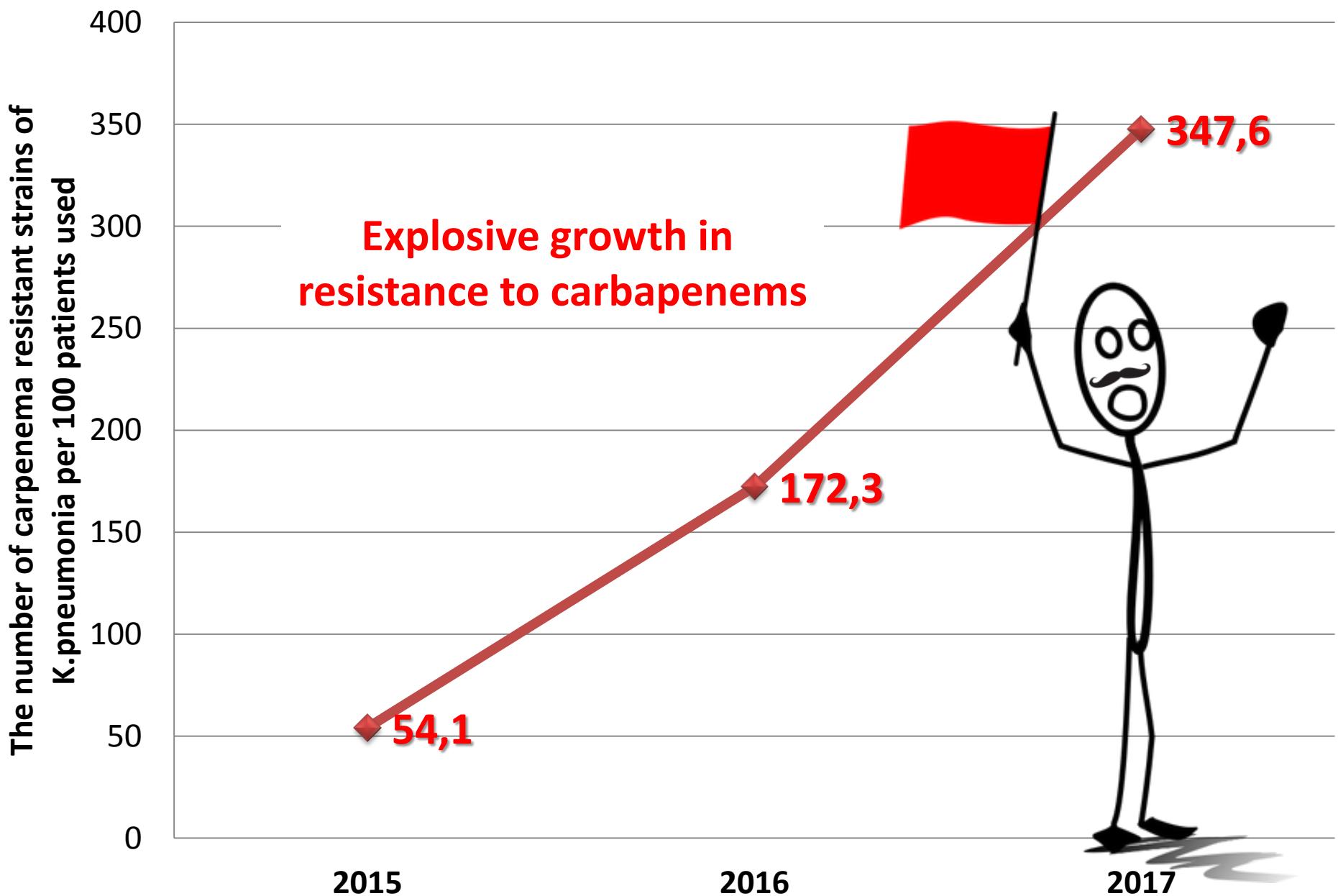
Cases of microbial isolation



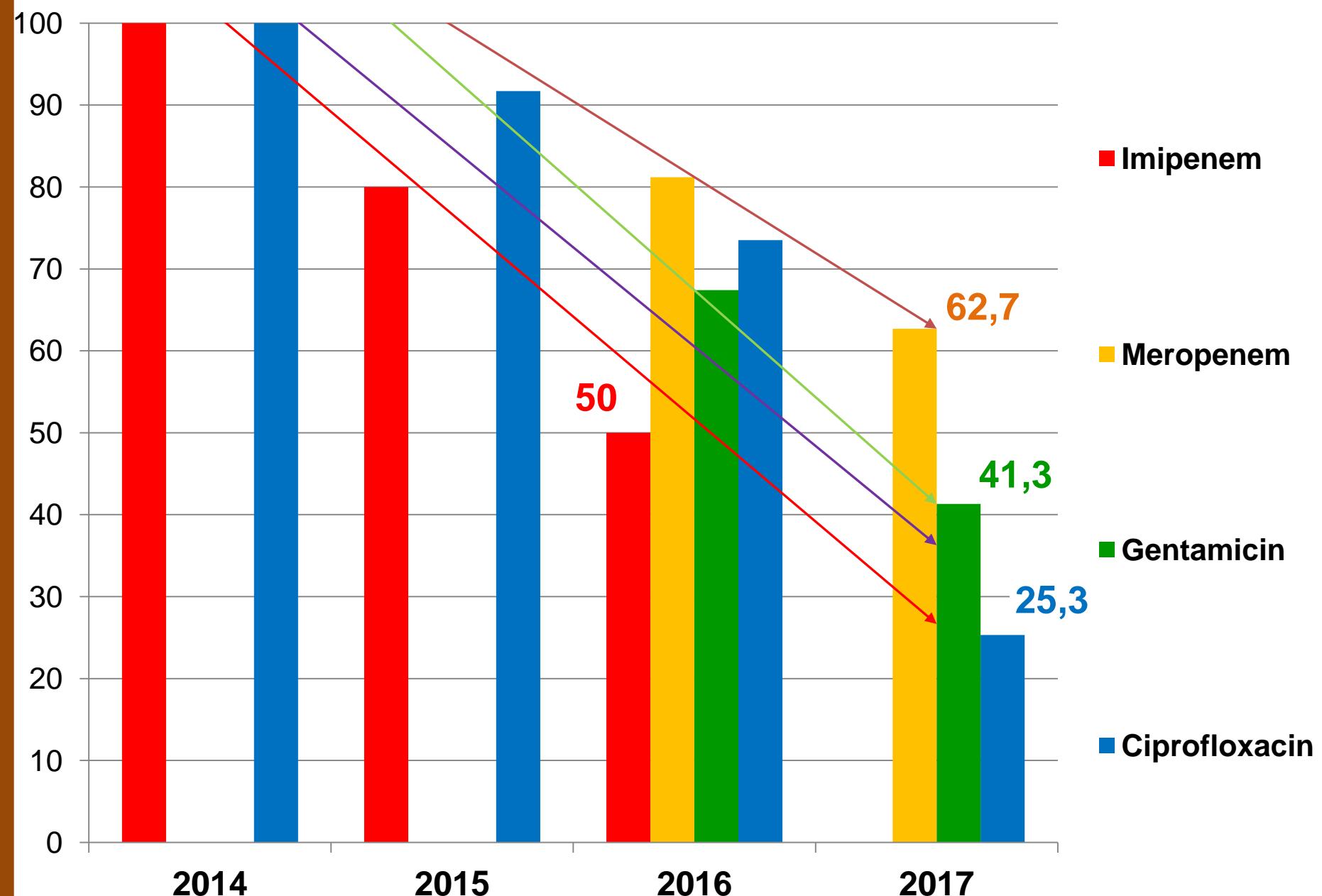
Long-term dynamics of changes in the structure of microorganisms isolated from all types of biological material (2015-2017)



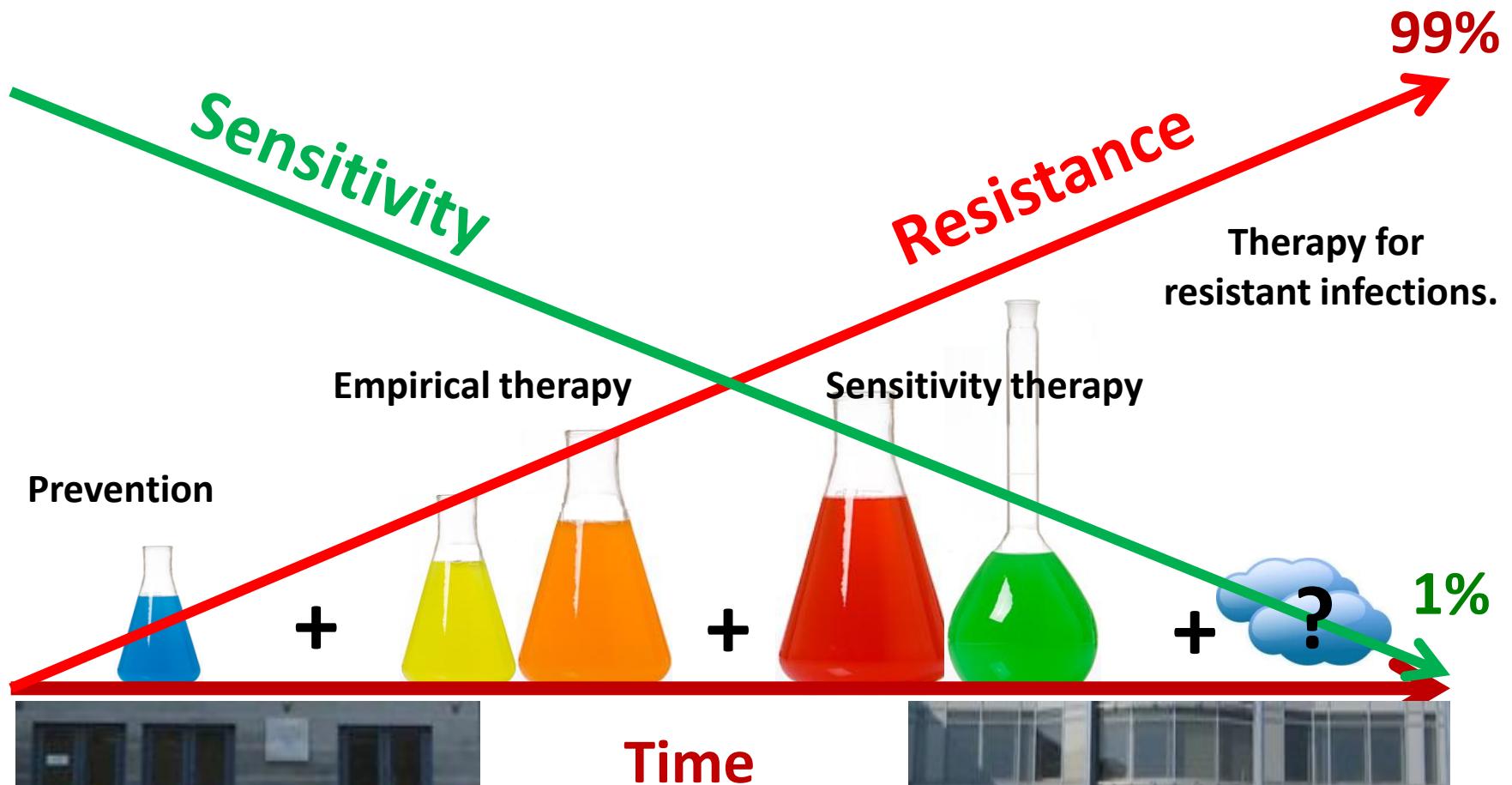
Changes in the resistance of K.pneumonia strains isolated from patients (2015-2017)



Sensitivity of Klebsiella pneumoniae strains isolated from stool against CARBA, GENTA, CIPRO 2014-2017



DEPARTMENT WITH ABSOLUTE RESISTANCE



Patient characteristics

Colonization of Klebsiella pneumoniae after BMT

n=12

| | | |
|---------------------------|-----------|-----|
| Возраст | 36(22/62) | |
| Диагноз | n | % |
| ОЛЛ | 3 | 25 |
| ХМЛ | 3 | 25 |
| НХЛ | 1 | 8 |
| ОМЛ | 5 | 42 |
| Вид ТГСК | | |
| а.н. | 10 | 83 |
| а.р. | 1 | 8 |
| гапло | 1 | 8 |
| Кондиционирование | | |
| Флюдарабин+Бусульфан | 12 | 100 |
| Профилактика РТПХ | | |
| Такролимус+ММФ | 2 | 17 |
| Бендомустин+Сиролимус+ | | |
| Руксолитиниб | 1 | 8 |
| Циклофосфан+Такролимус | 1 | 8 |
| Циклофосфан | 1 | 8 |
| Циклофосфан+Такролимус+МФ | 7 | 58 |

Inclusion criteria

- > 18 years old
- HSCT

Study days

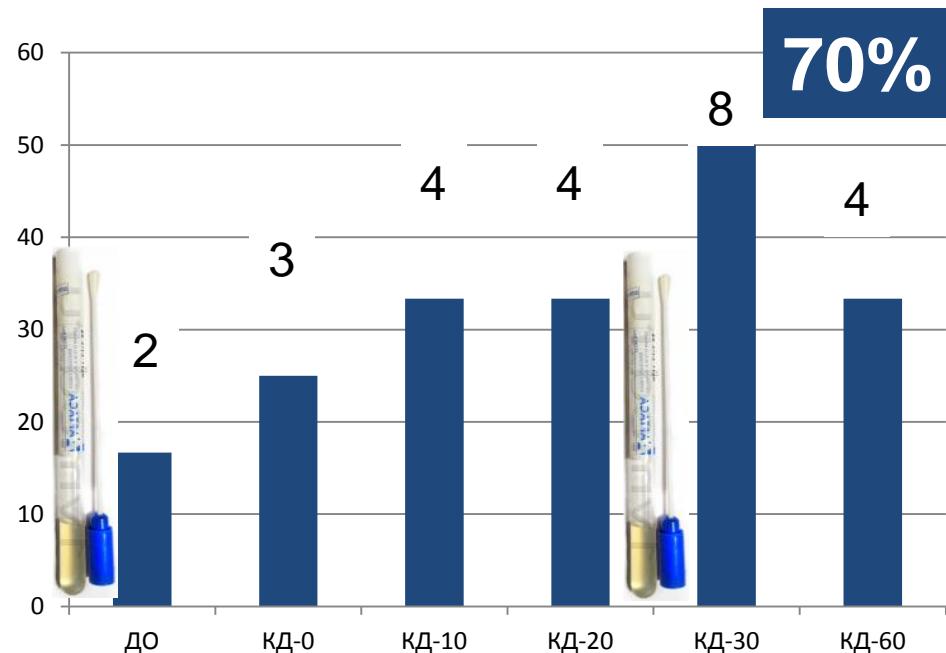
- 1- Day of admission to the clinic
- 2-D-0-
- 3-D + 10
- 4-D + 20
- 5-D + 30
- 6-D + 60

Research methods

- 1-Microbiological method
- 2-sensitivity to a/b
- 3-Detection of ESBL CES, Carbapenem KEC, ESBL E. coli, Carbapenem E. coli, VRE using chromogenic media CHROMagar

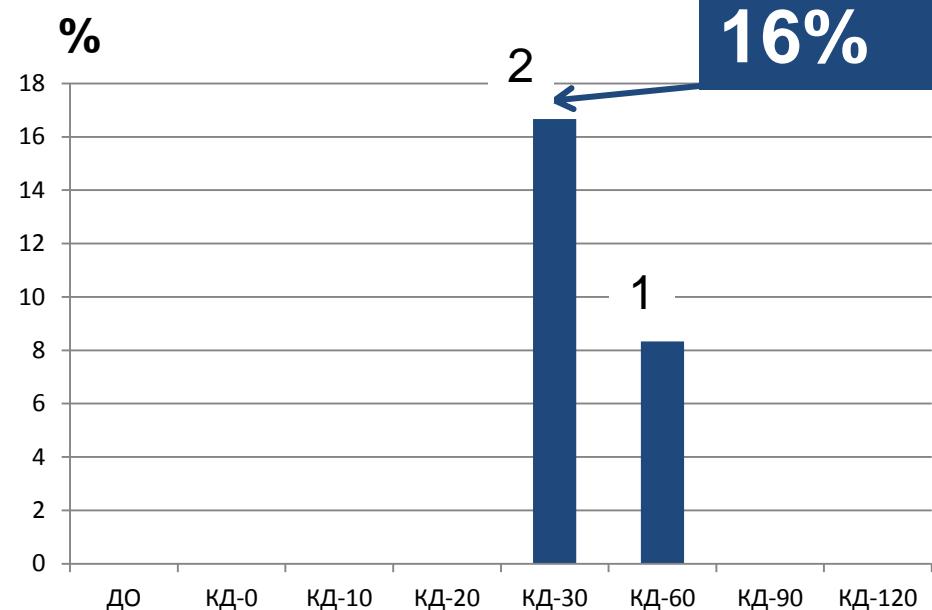
Feces

Klebsiella pneumoniae



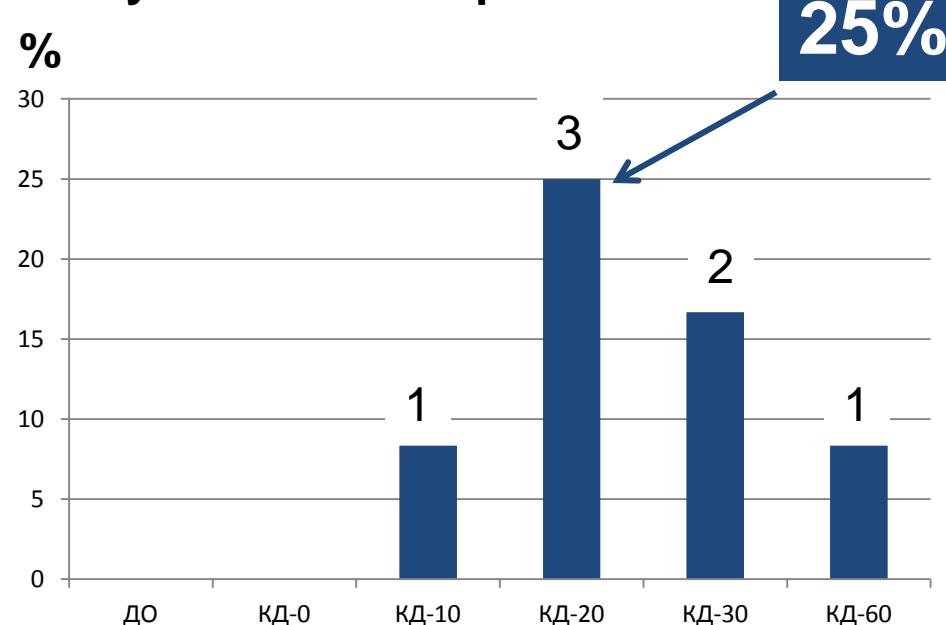
Urine

Klebsiella pneumoniae

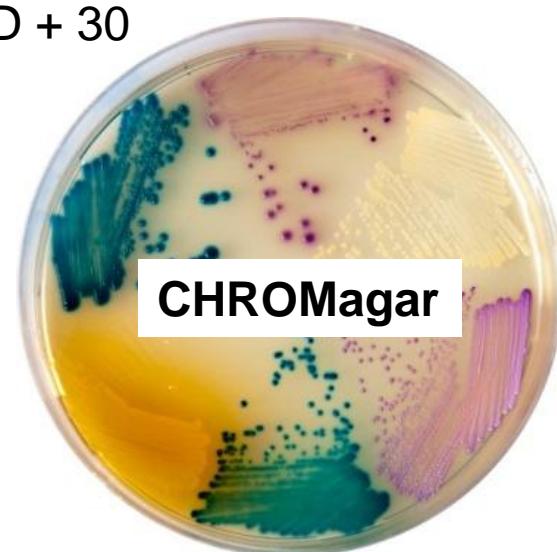


Pharynx

Klebsiella pneumoniae



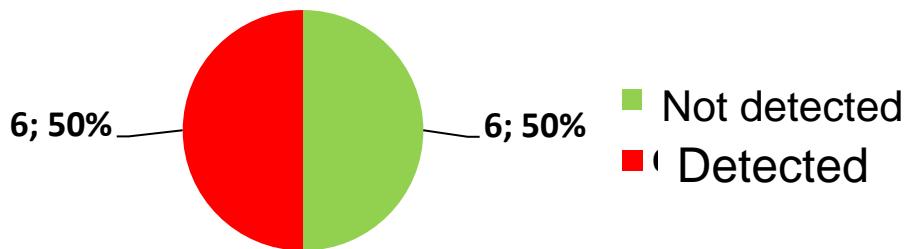
1- Day of admission to the clinic
2-D + 30



Chromogenic environment CHROMagar to identify

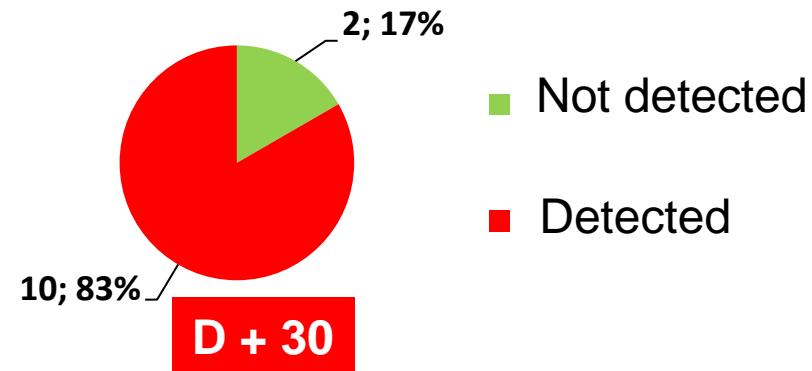
**ESBL KEC, Carbapenem KEC, ESBL E.coli,
Carbapenem E.coli, VRE (faecium, faecalis)**

Before BMT

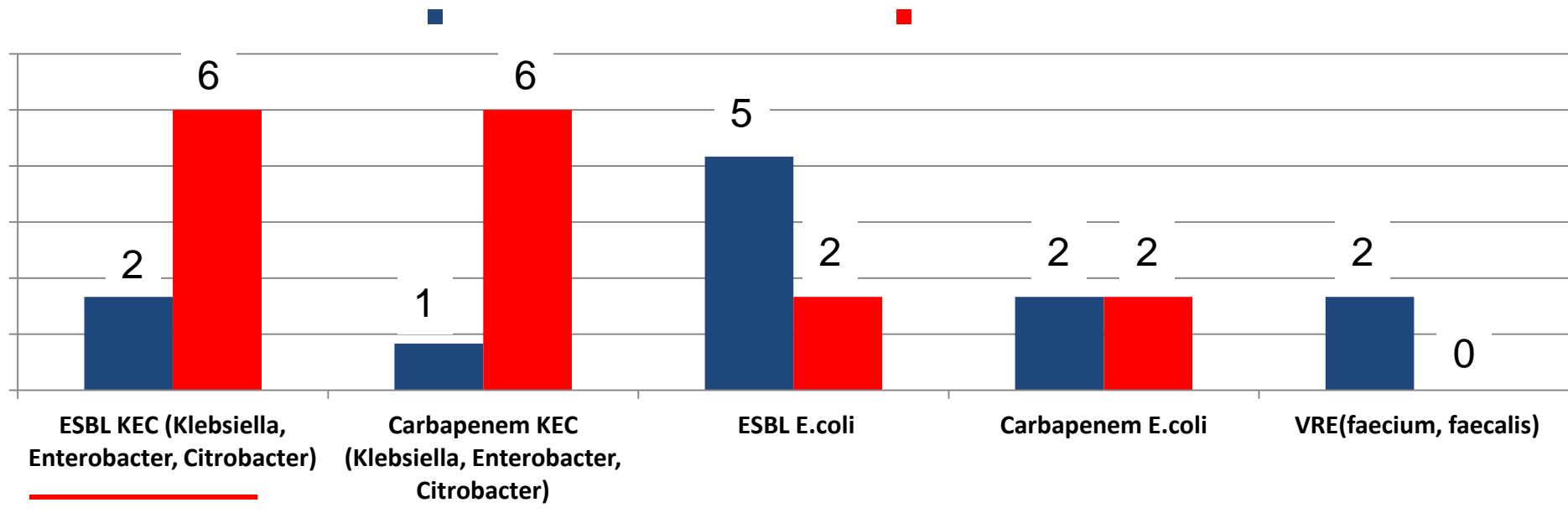


Before BMT

After BMT D + 30



D + 30

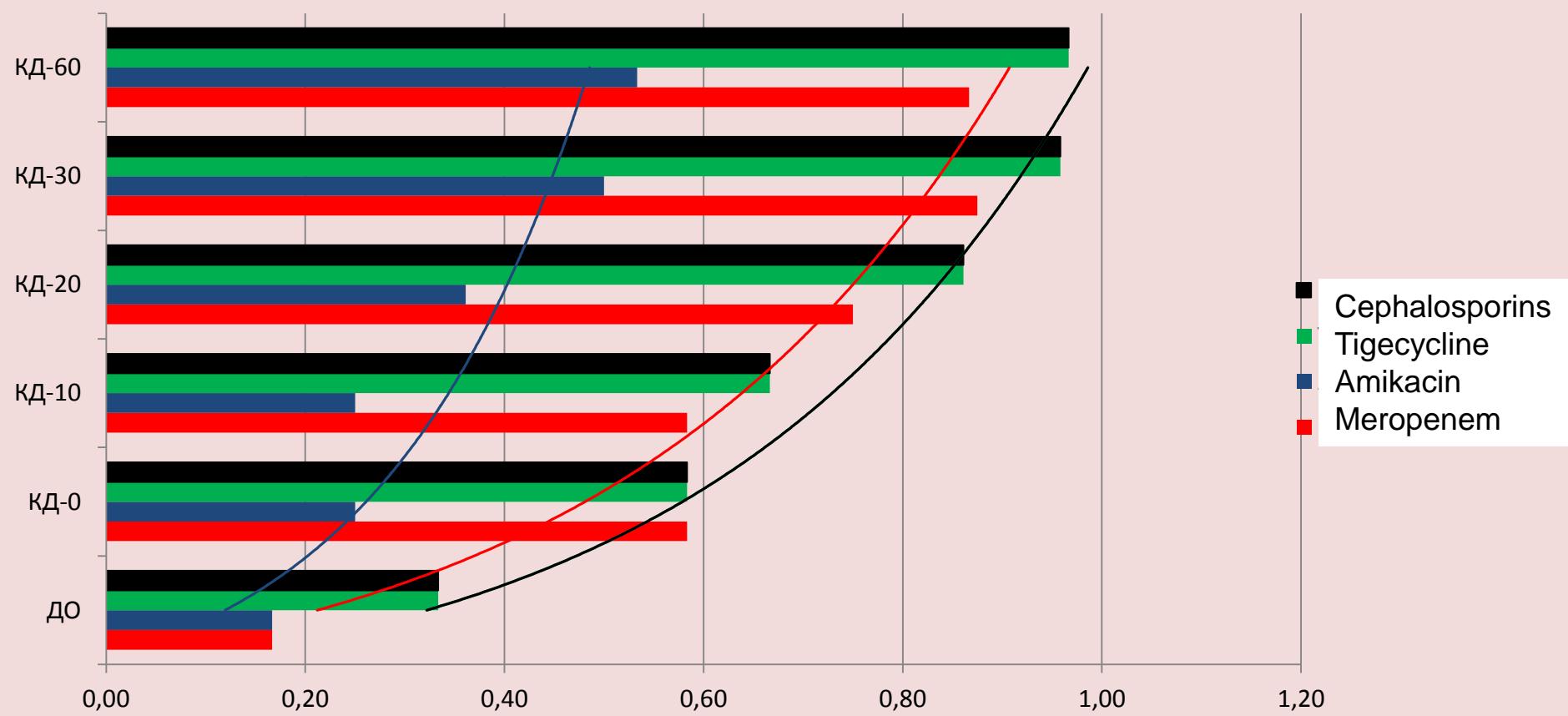




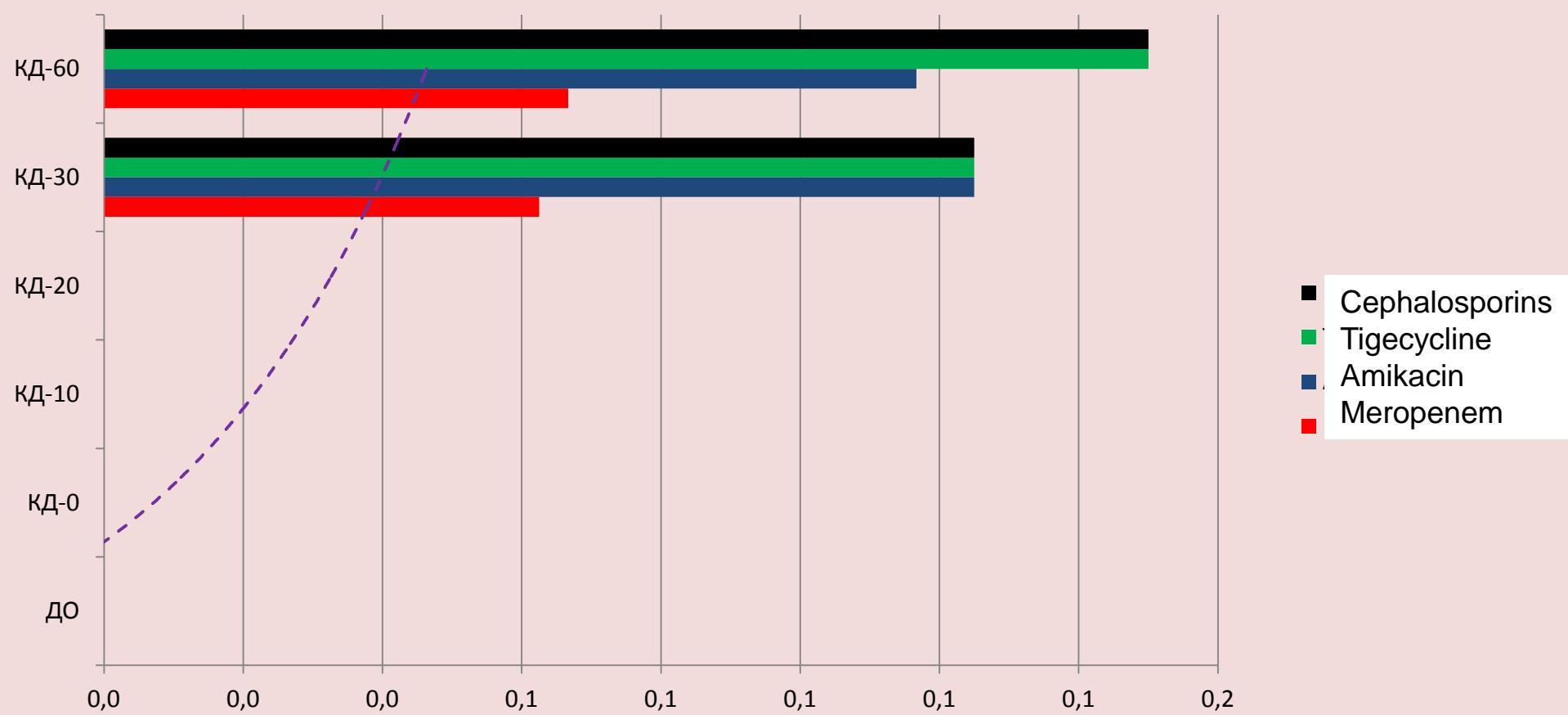
Feces

Increased colonization, increased antibiotic resistance

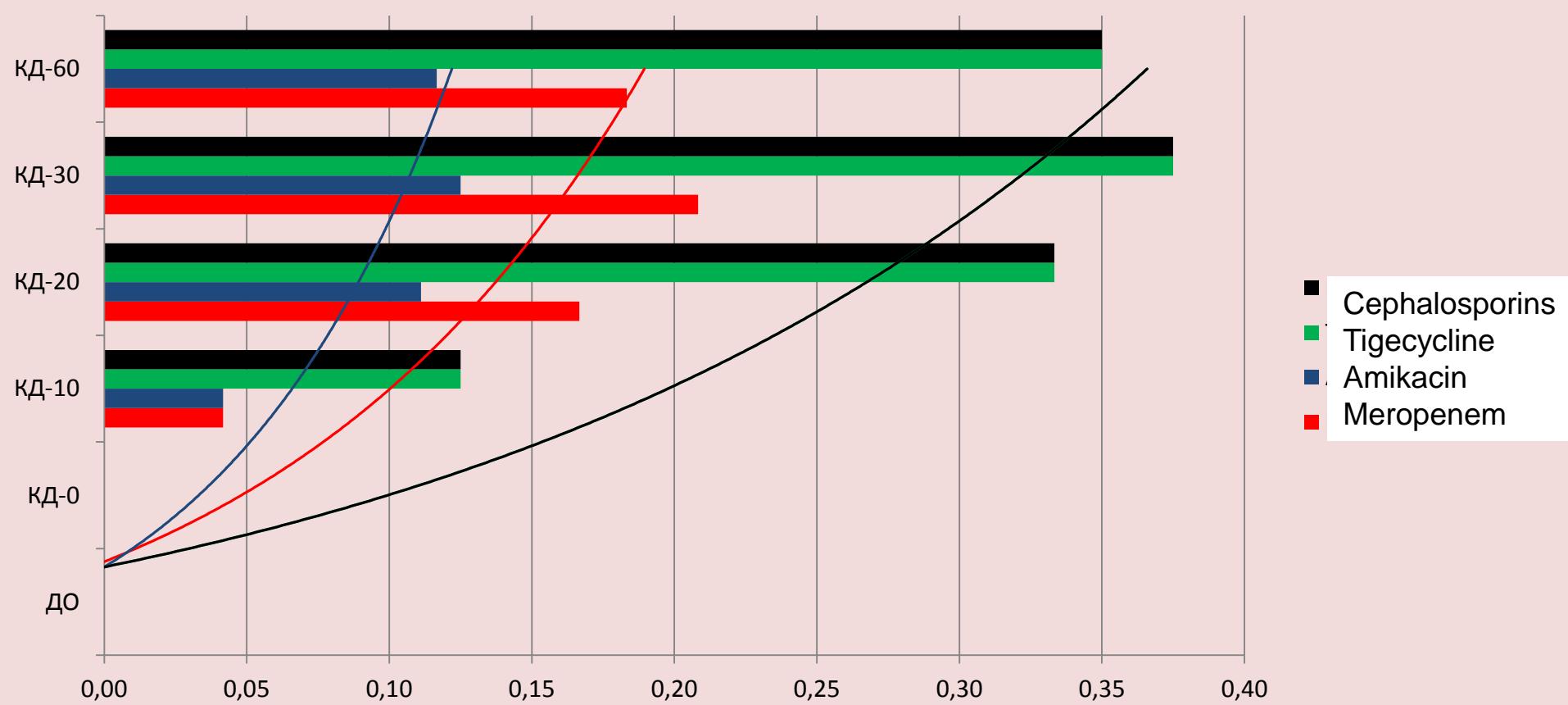
n=12 patients after BMT



n=12 patients after BMT



n=12 patients after BMT



Resistance Acinetobacter spp. in the hematology department (HD)

intensive care unit (ICU)

| 55 isolates | HD/ICU | HD/ICU | HD/ICU |
|-------------------------------|---------------|--------------|---------------|
| | S% | I% | R% |
| Cefoperazone/Sulbactam | ★82/54 | 9/0 | 9/46 |
| (😢😢) Ciprofloxacin | 10/8 | 14/0 | 76/92 |
| (😢) Imipenem | 26/8 | 15/0 | 59/92 |
| (😢) Meropenem | 32/0 | 17/18 | 51/82 |
| (😢) Netilmicin | 34/0 | 5/0 | 61/100 |
| Tigecycline ★★ | 98/100 | 2/0 | 0/0 |

Recommendations for starting therapy **in the hematology department (HD)**
Cefoperazone/Sulbactam

Recommendations for starting therapy **in the intensive care unit (ICU)** **Tigecycline**

Resistance Klebsiella spp. in the hematology department (HD) intensive care unit (ICU)

| 825 isolates | HD/ICU | HD/ICU | HD/ICU |
|-------------------------------|-----------------|--------------|--------------|
| | S% | I% | R% |
| Amikacin ☹ | 36/15 | 19/11 | 66/53 |
| Aztreonam ☹ | 7/2 | 0/1 | 93/97 |
| Cefoperazone/Sulbactam | ★63/27 ☹ | 2/2 | 35/71 |
| Ciprofloxacin ☹☹ | 9/5 | 2/6 | 89/89 |
| Colistin ★★ | 98/93 | 0/0 | 2/7. |
| Imipenem | ★76/37 ☹ | 1/4 | 23/59 |
| Meropenem | ★63/36 ☹ | 3/8 | 34/56 |
| Netilmicin ☹ | 33/13 | 6/0 | 61/87 |
| Tigecycline ★★ | 95/93 | 3/2 | 2/3. |

Recommendations for starting therapy in the hematology department (HD)

Recommendations for starting therapy in the intensive care unit (ICU)

Meropenem
Imipenem
Tigecycline
Colistin

Resistance Enterobacter spp. in the hematology department (HD) intensive care unit (ICU)

| 515 isolates | HD/ICU | HD/ICU | HD/ICU |
|-------------------------------|----------------|-------------|--------------|
| | S% | I% | R% |
| Amikacin | 94/79 | 1/8 | 5/13. |
| Aztreonam | 30/12 | 0/3 | 70/85 |
| Cefoperazone/Sulbactam | 97/74 | 1/10 | 2/16. |
| Ciprofloxacin | 17/25 | 6/25 | 77/50 |
| Colistin | 94/100 | 0/0 | 6/0. |
| Imipenem | 99/95 | 0/0 | 1/5. |
| Meropenem | 92/100 | 0/8 | 8/0. |
| Netilmicin | 59/13 | 8/0 | 33/87 |
| Tigecycline | 100/100 | 0/0 | 0/0. |

Recommendations for starting therapy **in the hematology department (HD)** **Cefoperazone/Sulbactam** **Amikacin** **Meropenem**

Recommendations for starting therapy **in the intensive care unit (ICU)** **Tigecycline** **Colistin**

Resistance E. Faecalis / E. Faecium in the hematology department (HD) intensive care unit (ICU)

| 515 isolates | HD/ICU | HD/ICU | HD/ICU |
|----------------------|-----------|--------|---------|
| E. faecalis | S% | I% | R% |
| Ampicillin | ★ 85/85 | 1/4 | 14/11. |
| Ampicillin/Sulbactam | ★ 95/96 | 0/0 | 5/4.. |
| Linezolid | ★ 100/100 | 0/0 | 0/0 |
| Tigecycline | ★ 100/96 | 0/0 | 0/4. |
| Vancomycin | ★ 100/96 | 0/0 | 0/4. |
| E. faecium | S% | I% | R% |
| Ampicillin | (?) 0/0 | 0/0 | 100/100 |
| Ampicillin/Sulb. | (?) 0/0 | 0/0 | 100/100 |
| Linezolid | ★ 100/100 | 0/0 | 0/0. |
| Tigecycline | ★ 100/100 | 0/0 | 0/0. |
| Vancomycin | ★ 92/75 | 0/0 | 8/25. |
| Clindamycin | (?) 0/0 | 0/0 | 100/100 |

| | HD/ICU | HD/ICU | HD/ICU |
|---------------------------|-----------|--------|--------|
| Staphylococcus | S% | I% | R% |
| Daptomycin | ★ 100/100 | 0/0 | 0/0. |
| Ampicillin/Sulbactam | (?) 18/17 | 0/0 | 82/83 |
| Linezolid | ★ 99/97 | 0/0 | 1/3. |
| Tigecycline | ★ 99/100 | 0/0 | 1/0. |
| Vancomycin | ★ 100/100 | 0/0 | 0/0. |
| Clindamycin | (?) 56/45 | 7/9 | 37/46 |
| Oxacillin | (?) 24/10 | 0/0 | 76/90 |
| Streptococcus spp. | S% | I% | R% |
| Levofloxacin | (?) 63/60 | 5/10 | 32/30 |
| Ampicillin/Sulb. | 67/50 | 3/5 | 30/45 |
| Ceftriaxone | (?) 35/36 | 18/11 | 47/53 |
| Tigecycline | ★ ★ 100 | 0 | 0. |
| Vancomycin | ★ ★ 90 | 0/0 | 10. |
| Clindamycin | (?) 52/20 | 3/10 | 45/70 |
| Erythromycin | (?) 18/10 | 10/0 | 72/90 |

The problem is drug combinations.

Mono or combination therapy?

Have you asked a clinical pharmacologist?

Who is prescribing more than two antibiotics?

Too many antibiotics!

Too expensive!

Well, who treats like that?

Resistant therapy Klebsiella spp.

Bacteremia *Klebsiella pneumoniae* KPC+
- effect only from the combined ABT

n=41

78% -внутрибольничная инфекция

1) 28-day mortality

- Mono: 57,8%
- Сombo: 13,3% p=0.01

2) Multivariate analysis:

Combined ABT was significantly associated with patient survival,
 $OR = 0.07$ (95% CI 0.009-0.71, $p = 0.02$)

The most effective combinations:

Tigecycline + Carbapenem

Colistin + Carbapenem

carbapenem-resistant strains

Combination therapy

Mortality

Колистин

+

Carbapenems

Tigecyclin

Fluoroquinolones

Tigecyclin

- +

Carbapenems

Aminoglycosides

Monotherapy

Colistin

Tigecycline

Carbapenems

Gentamicin

Ampicillin-sulbactam

Piperacillin-Tazobactam



Effect of combination therapy KPC Klebsiella spp. – with bloodstream infections

n=125

Mortality %

Mono - 54,3

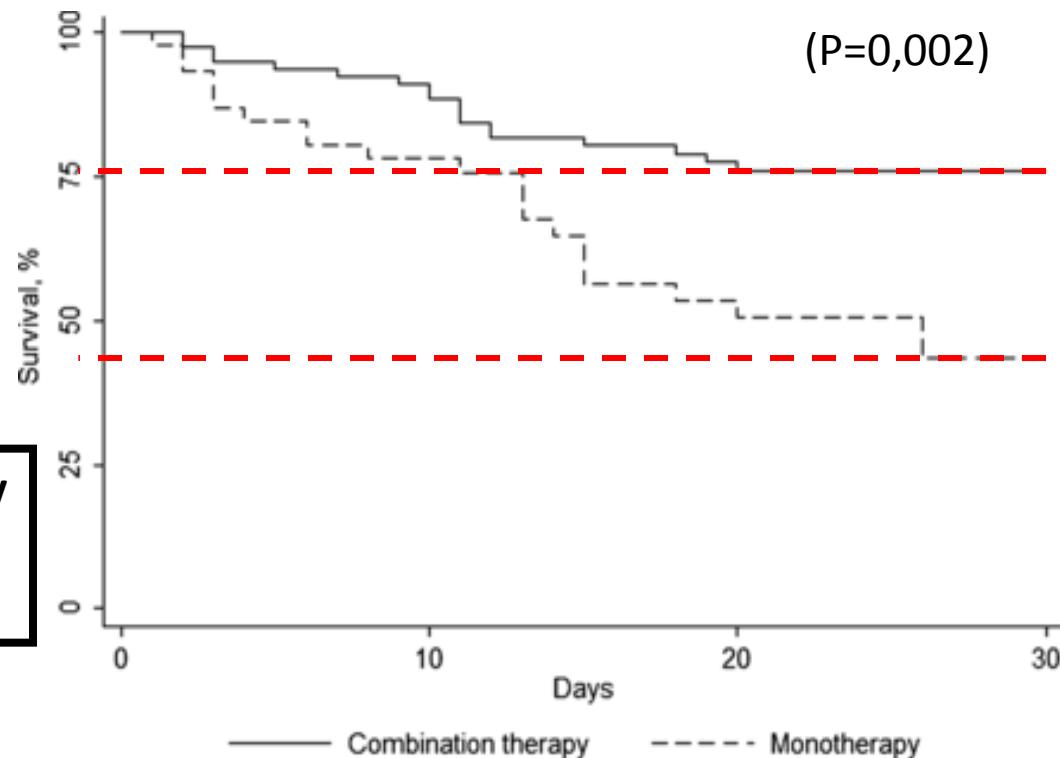
2 АБ - 41,1

3 АБ - 17,4

Inadequate initial antimicrobial therapy

- cause of death

(P = 0,003)



Tigecycline + Carbapenems + Colistin

P = 0,01

Tigecycline + Carbapenems + Gentamicin

Colistin + Carbapenems + Gentamicin

Resistant therapy Klebsiella spp.

N=250

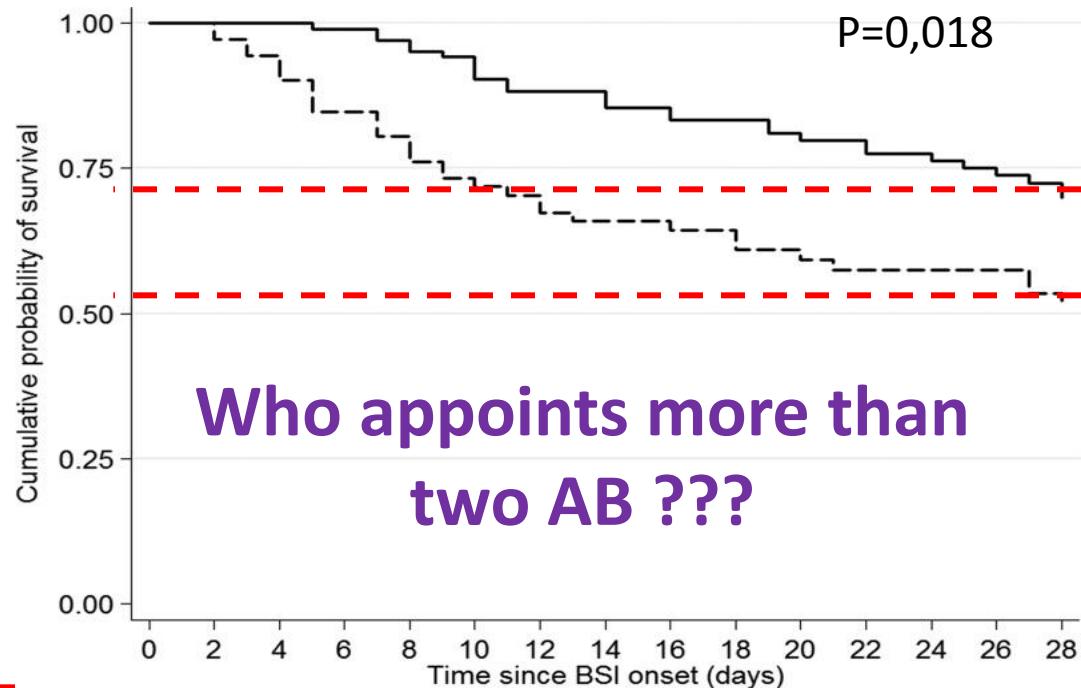
Mortality %

Mono: 44,4%

Combo: 27,2%

Monotherapy - Mortality %

| | |
|------------|------|
| Tigecyclin | 40.4 |
| Colistin | 54.5 |
| Carbapenem | 58.0 |



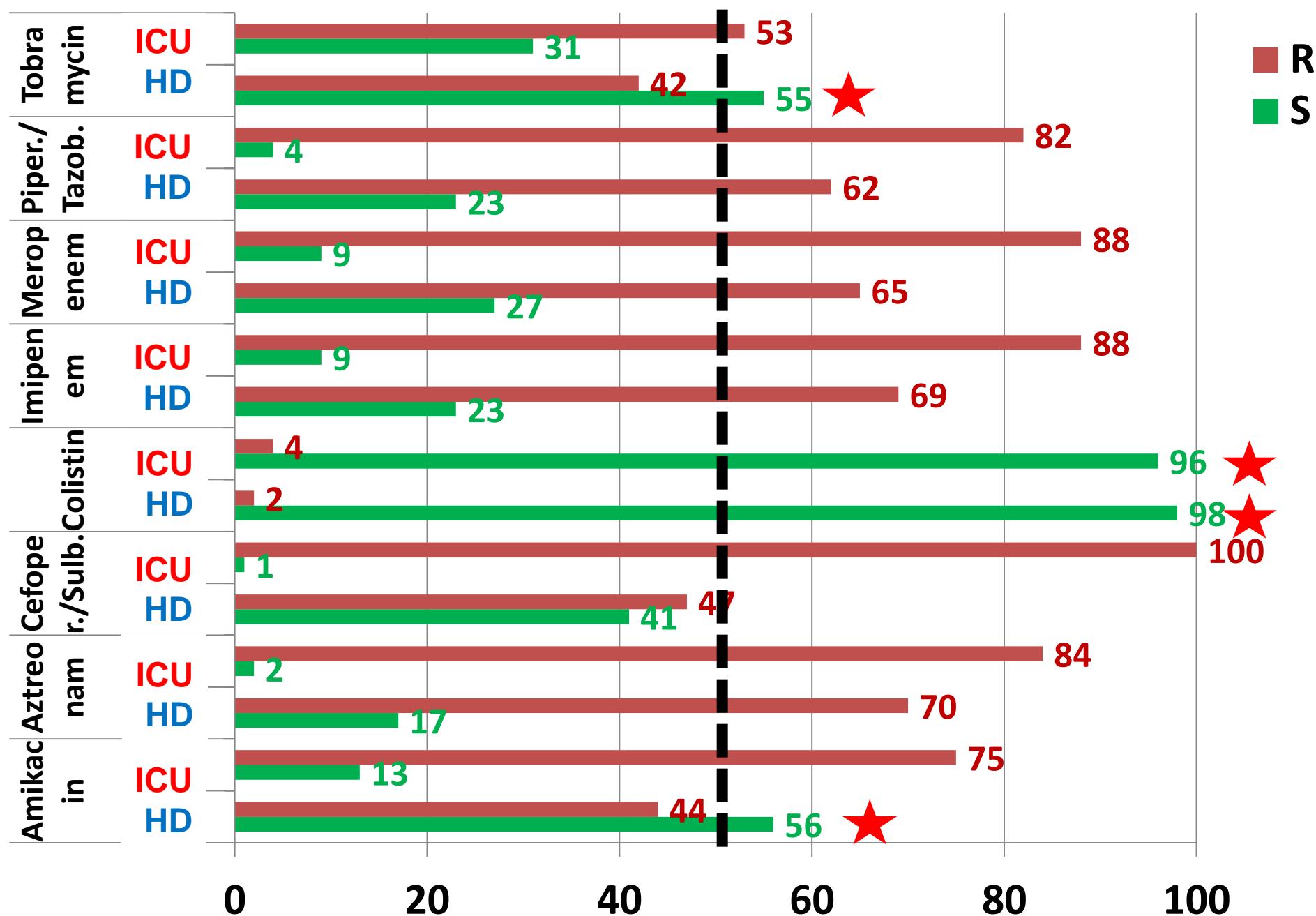
Who appoints more than
two AB ???

Combination Therapy

- Carbapenem + Tigecyclin + AH or Colistin
 - Tigecyclin + Colistin + AG
 - Tigecyclin + Colistin
- Mortality (%)
- | |
|-------|
| 0% |
| 27.3% |
| 23.8% |

Pseudomonas spp.

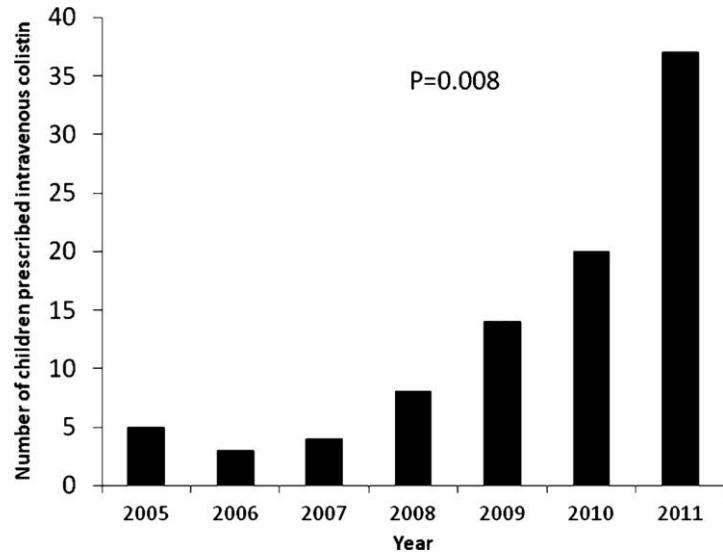


Pseudomonas aeruginosa

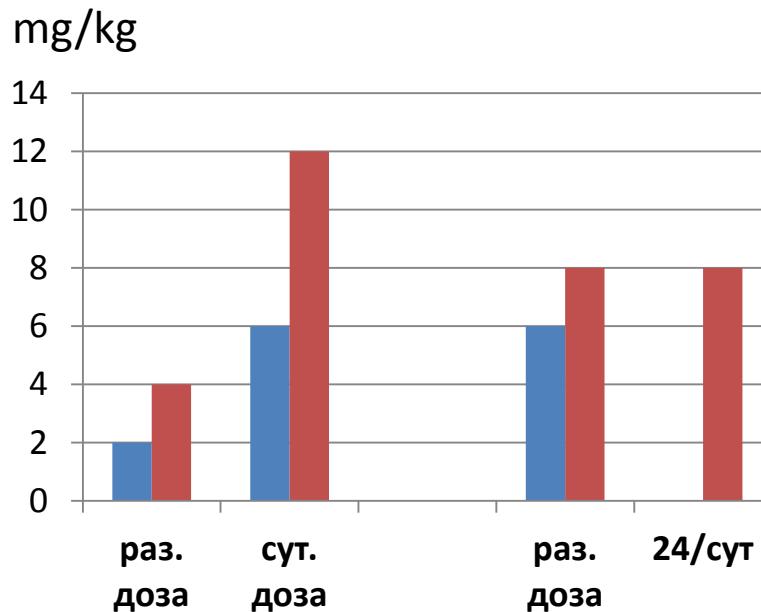
1950

COLISTINE =

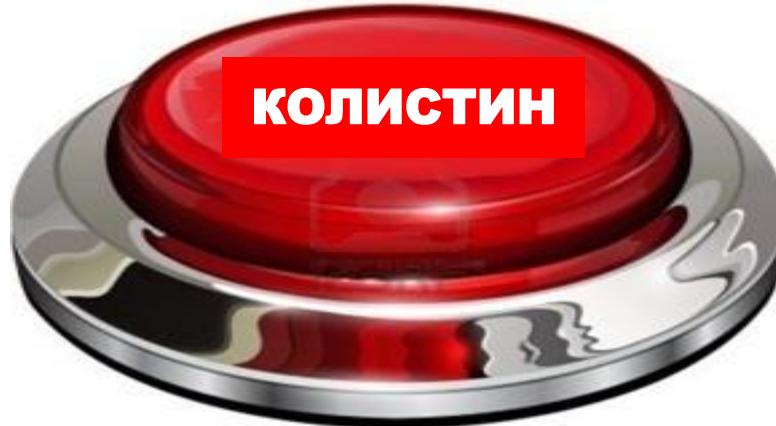
colistin A (polymyxin E1) +
colistin B (polymyxin E2)



Pediatr Infect Dis J. 2013 Jan; 32(1):17–22. Pranita D. Tamma
The Use of Intravenous Colistin Among Children in the United States



Colistin 9.000.000 Ui ev,
then 4.500.000 x 2 ev



Colistin problems

- 1-resistance
- 2 is not applicable in monotherapy
- 3-limit nephrotoxicity



4 medicines left ???

TIENAM

+ TIGACYCLINE

+ COLISTINE



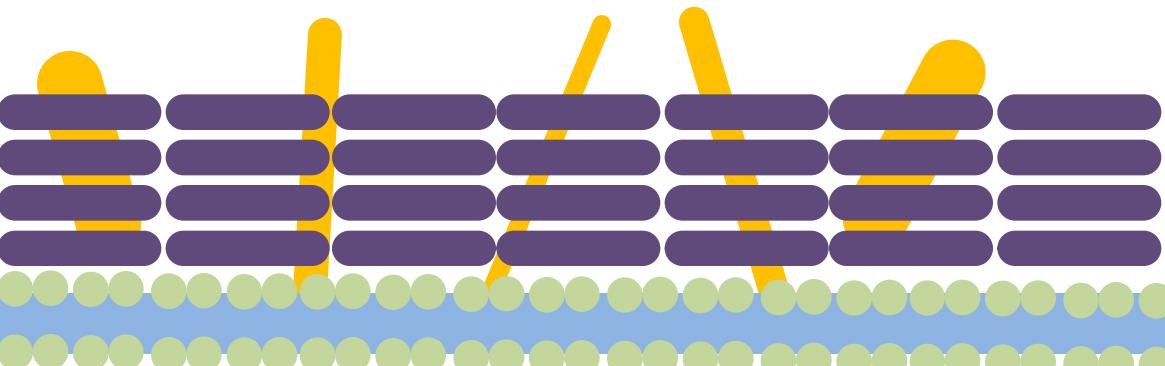
+ AMEN

Mechanisms of resistance to carbapenems

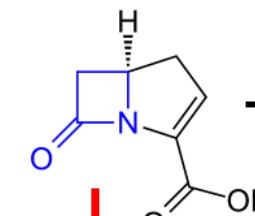
Teichoic Acids

Peptidoglycans

Cytoplasmic
membrane

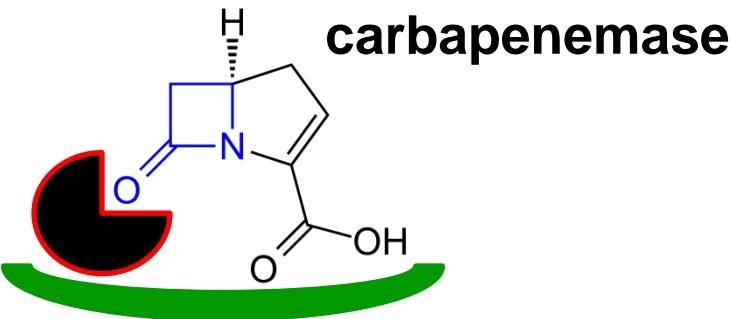
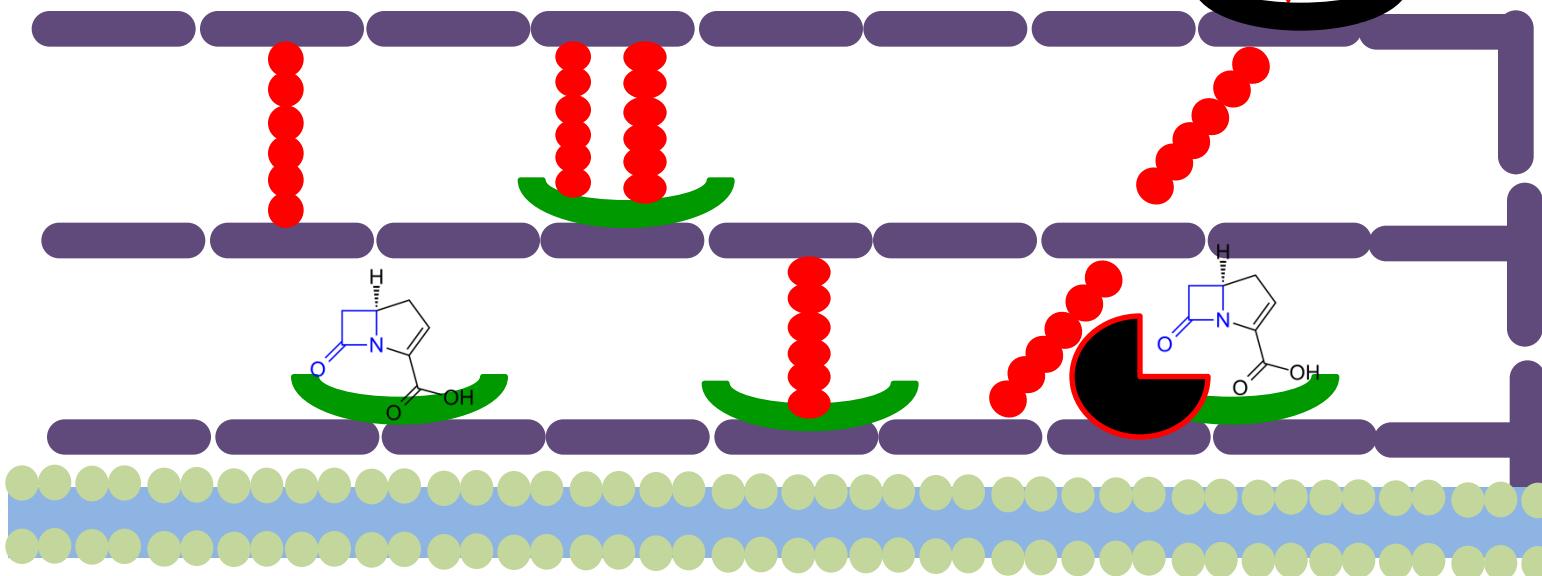


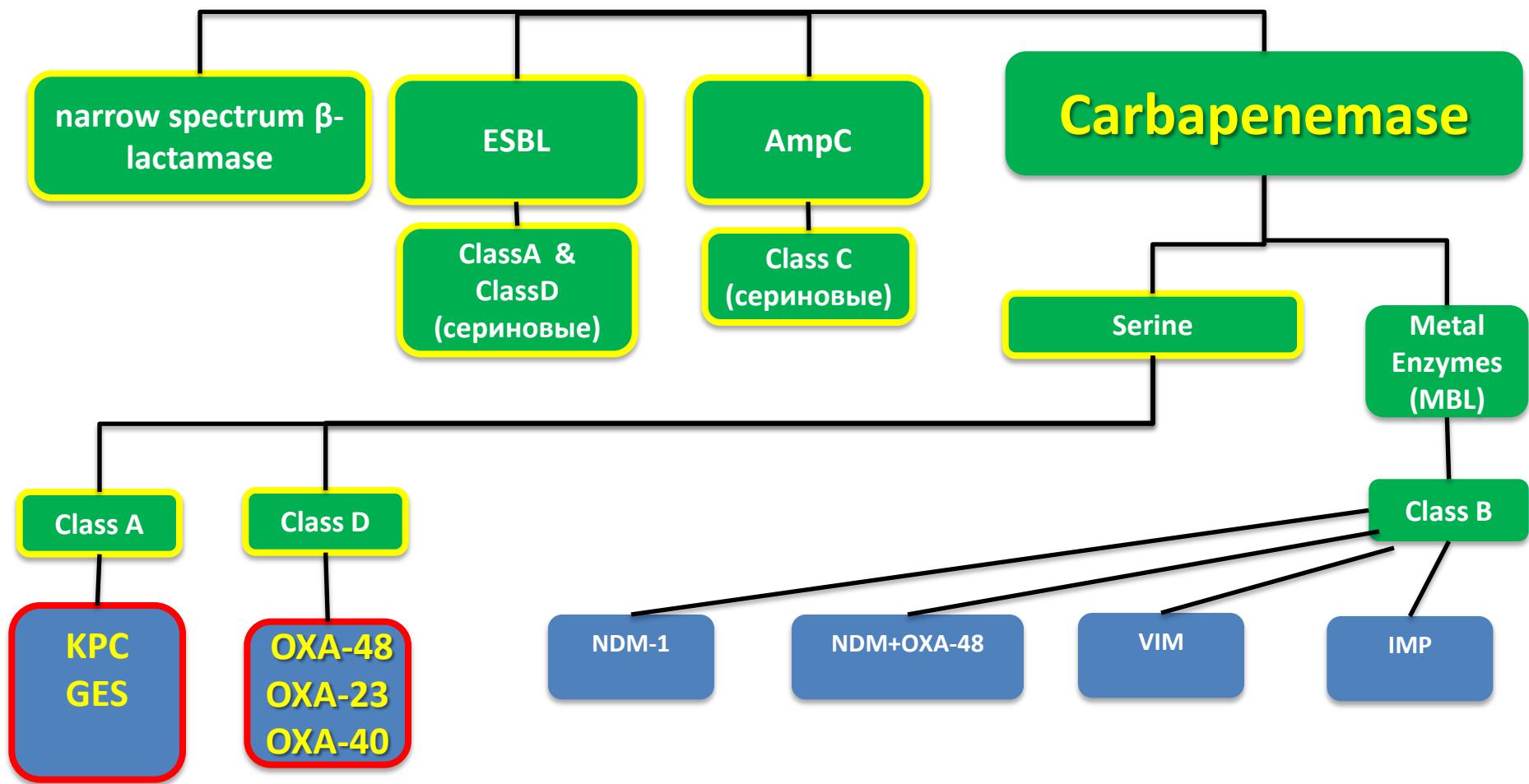
Transpeptidase



Tightness

Removal

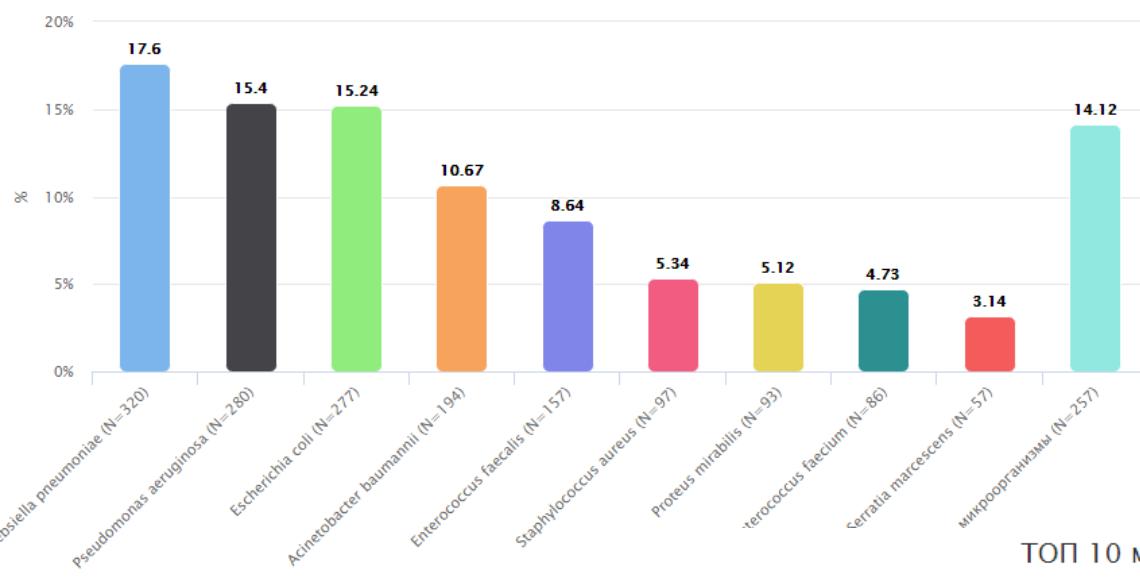




St. Petersburg

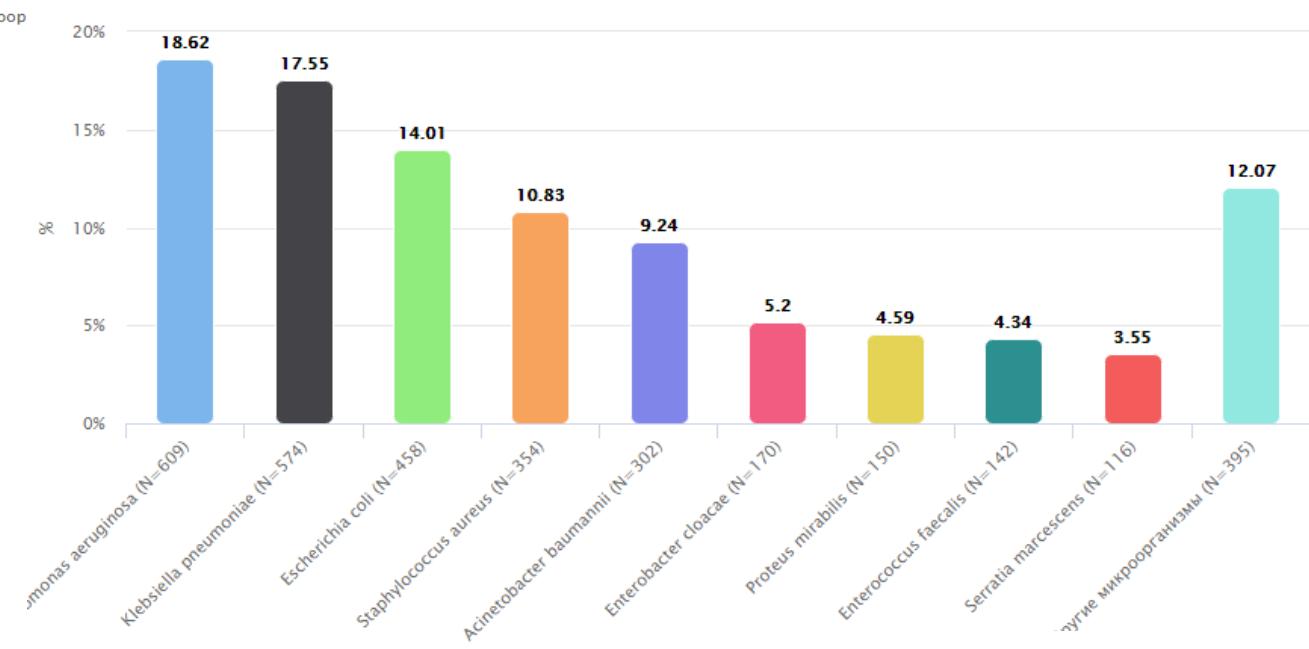
Klebsiella spp

ТОП 10 микроорганизмов (N=1818)



MOSCOW Pseudomonas spp.

ТОП 10 микроорганизмов (N=3270)



Микроорганизмы

МАКМА

16.11.2018

Carbapenemase - Klebsiella pneumoniae

<http://www.amrmap.ru>

St. Petersburg

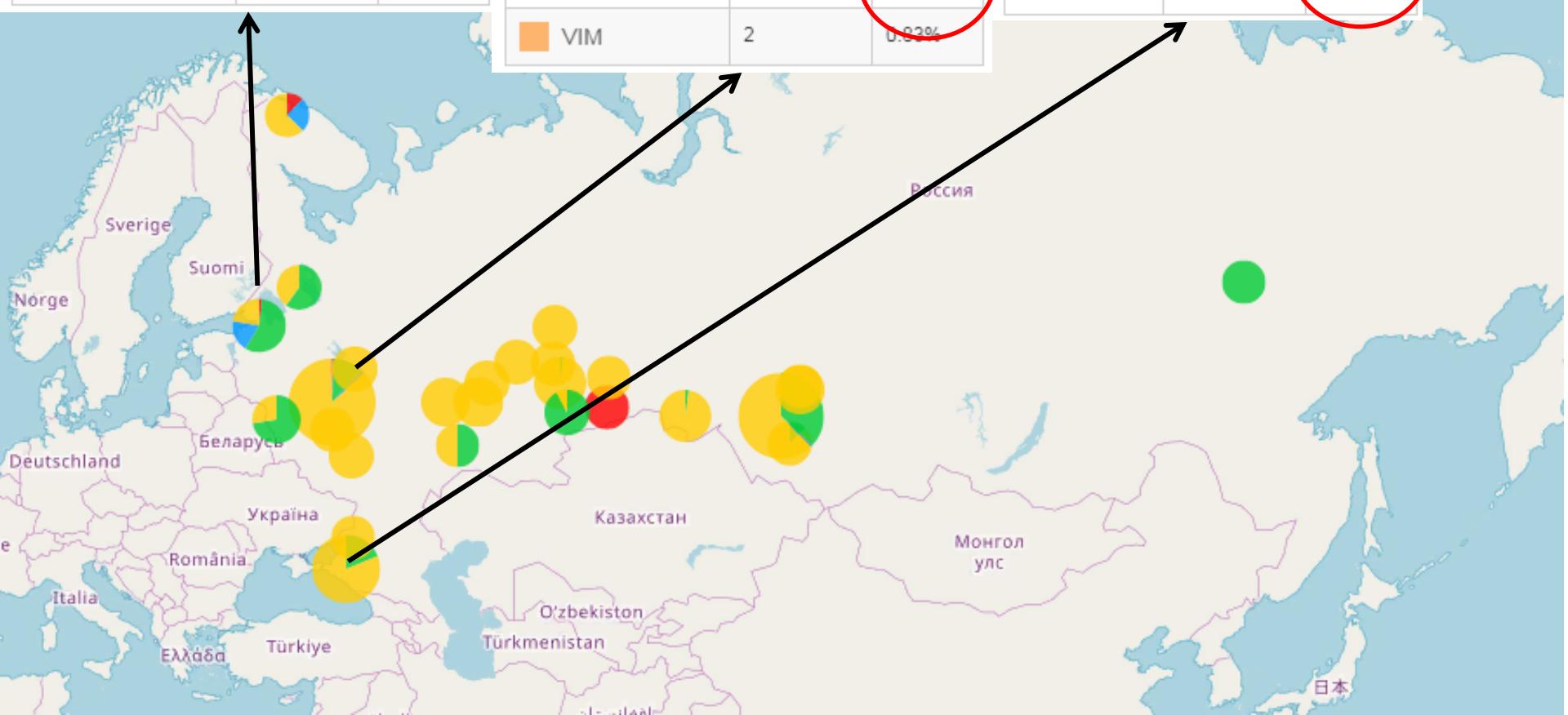
| Категория | Количество | Процент |
|----------------|------------|---------|
| KPC | 1 | 1.72% |
| NDM | 33 | 56.9% |
| NDM+OXA-48-гр. | 11 | 18.07% |
| OXA-48-гр. | 13 | 22.41% |

Moscow

| Категория | Количество | Процент |
|----------------|------------|---------|
| KPC | 1 | 0.42% |
| NDM | 27 | 11.25% |
| NDM+OXA-48-гр. | 4 | 1.67% |
| OXA-48-гр. | 206 | 85.83% |
| VIM | 2 | 0.02% |

Krasnodar

| Категория | Количество | Процент |
|------------|------------|---------|
| NDM | 26 | 18.84% |
| OXA-48-гр. | 112 | 81.16% |



Genetic markers of antibiotic resistance



| | | | | Carbapenemas e+/- | Multiplex PCR(NDM,OXA- 48,KPC)_Carba- penemase type | | |
|----|---------|------------|--------------------|----------------------|--|------------|------------|
| 1 | 4751803 | 12.03.2018 | моча | KL.pneumoniae | 12.03.2018 | NDM+OXA-48 | NDM+OXA-48 |
| 2 | 4811833 | 03.04.2018 | ЦВК | Acinet. Sp | 03.04.2018 | отр | отр |
| 3 | 4877268 | 28.04.2018 | кал | E.coli | 28.04.2018 | отр | отр |
| 4 | 4842587 | 14.04.2018 | моча | KL.pneumoniae | 14.04.2018 | отр | отр |
| 5 | 4811458 | 06.04.2018 | кал | KL.pneumoniae | 06.04.2018 | отр | отр |
| 6 | 4811694 | 10.04.2018 | катетер | Pseud. sp | 10.04.2018 | отр | отр |
| 7 | 4812047 | 04.04.2018 | кал | E.coli | 04.04.2018 | NDM | NDM |
| 8 | 4900878 | 10.05.2018 | мочевой катетер | KL.pneumoniae | 10.05.2018 | KPC | KPC |
| 9 | 4798614 | 03.04.2018 | БАЛ | KL.pneumoniae | 03.04.2018 | KPC | KPC |
| 10 | 4812183 | 04.04.2018 | миндалины | KL.pneumoniae | 04.04.2018 | NDM+OXA-48 | NDM+OXA-48 |
| 11 | 4877702 | 25.04.2018 | промывные воды бро | Ps.sp | 25.04.2018 | отр | отр |
| 12 | 4812183 | 04.04.2018 | миндалины | Ps.sp | 04.04.2018 | KPC | KPC |
| 13 | 4812184 | 04.04.2018 | кал | KL.pneumoniae | 04.04.2018 | KPC | KPC |
| 14 | 4707690 | 12.02.2018 | миндалины | KL.pneumoniae | 12.02.2018 | KPC+NDM | KPC+NDM |
| 15 | 4707665 | 12.02.2018 | ЦВК | KL.pneumoniae | 12.02.2018 | KPC+NDM | KPC+NDM |
| 16 | 4713974 | 19.02.2018 | моча | KL.pneumoniae | 19.02.2018 | отр | отр |
| 17 | 4713251 | 16.02.2018 | ЦВК | KL.pneumoniae | 16.02.2018 | OXA-48 | OXA-48 |
| 18 | 4713086 | 15.02.2018 | зев | KL.pneumoniae | 15.02.2018 | NDM+OXA-48 | NDM+OXA-48 |
| 19 | 4712994 | 15.02.2018 | кровь | KL.pneumoniae | 15.02.2018 | отр | отр |
| 20 | 4697973 | 12.02.2018 | моча | KL.pneumoniae | 12.02.2018 | KPC | KPC |
| 21 | 4616198 | 27.12.2017 | кал | KL.pneumoniae | 27.12.2017 | NDM | NDM |
| 22 | 4697983 | 12.02.2018 | кровь | KL.pneumoniae | 12.02.2018 | NDM | NDM |
| 23 | 4707798 | 12.02.2018 | пром. Воды бронхов | KL.pneumoniae | 12.02.2018 | KPC+NDM | KPC+NDM |
| 24 | 4707924 | 13.02.2018 | БАЛ | KL.pneumoniae | 13.02.2018 | NDM | NDM |
| 25 | 4707916 | 13.02.2018 | моча | KL.pneumoniae | 13.02.2018 | отр | отр |
| 26 | 4643601 | 12.01.2018 | кал | KL.pneumoniae | 12.01.2018 | отр | отр |
| 27 | 4643603 | 12.01.2018 | моча | KL.pneumoniae | 12.01.2018 | NDM | NDM |
| 28 | 4643513 | 11.01.2018 | половые органы | KL.pneumoniae | 11.01.2018 | NDM | NDM |
| 29 | 4646196 | 29.12.2017 | зев | KL.pneumoniae | 29.12.2017 | отр | отр |

Carbapenemase-Producing Enterobacteriaceae and Nonfermentative Bacteria, the Philippines, 2013–2016

John Mark Velasco, Maria Theresa Valderama, Trent Peacock,¹ Nirdnoy Warawadee, Kathleen Nogrado, Fatima Claire Navarro, Domingo Chua, Jr., Srijan Apichai, Ruekit Sirigade, Louis R. Macareo, Brett Swierczewski

Technical Appendix Table. Molecular resistance mechanisms of carbapenem-resistant clinical ($n = 45$) and environmental isolates ($n = 3$) from a tertiary-care military hospital in Manila, the Philippines, August 2013–April 2016*

| Identification | Source | Organism | Carba NP | bla _{NDM} | bla _{KPC} | bla _{VIM} | Month | Year | Imipenem MIC (μg/mL) | Meropenem MIC (μg/mL) | Sex | Hospital ward |
|------------------|-------------------|--------------------------|----------|--------------------|--------------------|--------------------|-------|------|----------------------|-----------------------|-----|-----------------------|
| Patient isolates | | | | | | | | | | | | |
| PH-0138-14 | Blood | <i>K. pneumoniae</i> | Pos | Pos | Neg | ND | Nov | 2013 | >8 | >8 | M | Neonatal ICU |
| PH-0542-14 | Soft tissue | <i>E. coli</i> | Pos | Pos | Neg | ND | Jun | 2014 | >8 | >8 | M | Pediatric |
| PH-0630-14 | Wound | <i>K. pneumoniae</i> | Pos | Pos | Neg | ND | Jul | 2014 | >8 | >8 | F | Female Medical |
| PH-0631-14 | Blood | <i>C. freundii</i> | Pos | Pos | Neg | ND | Jul | 2014 | >8 | >8 | M | Neurosurgery |
| PH-0756-14 | Catheter | <i>K. pneumoniae</i> | Pos | Pos | Neg | ND | Aug | 2014 | >8 | >8 | F | Medical ICU |
| PH-0787-14 | Endotracheal tip | <i>K. pneumoniae</i> | Pos | Pos | Neg | ND | Aug | 2014 | >8 | >8 | M | Medical ICU |
| PH-0837-14 | Catheter | <i>K. pneumoniae</i> | Pos | Pos | Neg | ND | Sep | 2014 | >8 | >8 | F | Female Surgical |
| PH-0846-14 | Catheter | <i>E. cloacae</i> | Pos | Pos | Neg | ND | Sep | 2014 | >8 | >8 | M | Male Medical Oncology |
| PH-0850-14 | Urine | <i>K. pneumoniae</i> | Pos | Pos | Neg | ND | Sep | 2014 | >8 | >8 | F | Female Medical |
| PH-0873-14 | Urine | <i>K. pneumoniae</i> | Pos | Pos | Neg | ND | Oct | 2014 | >8 | >8 | M | Surgical ICU |
| PH-0874-14 | Wound | <i>Klebsiella sp.</i> | Pos | Pos | Neg | ND | Oct | 2014 | >8 | >8 | M | Female Medical |
| PH-0901-14 | Wound | <i>C. freundii</i> | Pos | Pos | Neg | ND | Oct | 2014 | >8 | >8 | F | Female Surgical |
| PH-1037-14 | Catheter | <i>K. pneumoniae</i> | Pos | Pos | Neg | ND | Dec | 2014 | >8 | >8 | F | Female Medical |
| PH-1076-14 | Urine | <i>K. pneumoniae</i> | Pos | Pos | Neg | ND | Dec | 2014 | >8 | >8 | M | Medical ICU |
| PH-1078-14 | Blood | <i>K. pneumoniae</i> | Pos | Pos | Neg | ND | Dec | 2014 | >8 | >8 | F | Female Medical |
| PH-1088-14 | Endotracheal tip | <i>K. pneumoniae</i> | Pos | Pos | Neg | ND | Jan | 2015 | >8 | >8 | F | Medical ICU |
| PH-1093-14 | Tracheal aspirate | <i>K. pneumoniae</i> | Pos | Pos | Neg | ND | Dec | 2014 | >8 | >8 | F | Female Medical |
| PH-1099-14 | Endotracheal tip | <i>Acinetobacter sp.</i> | Pos | Pos | Neg | ND | Dec | 2014 | * | >8 | M | Surgical ICU |
| PH-1115-14 | Blood | <i>K. pneumoniae</i> | Pos | Pos | Neg | ND | Dec | 2014 | >8 | >8 | F | Female Medical |
| PH-1142-15 | Blood | <i>K. pneumoniae</i> | Pos | Pos | Neg | ND | Jan | 2015 | >8 | >8 | F | Female Medical |
| PH-1143-15 | Wound | <i>K. pneumoniae</i> | Pos | Pos | Neg | ND | Jan | 2015 | >8 | >8 | M | Neurology |
| PH-1150-15 | Catheter | <i>E. cloacae</i> | Pos | Pos | Neg | ND | Jan | 2015 | >8 | >8 | M | Neurosurgery |
| PH-1159-15 | Catheter | <i>K. pneumoniae</i> | Pos | Pos | Neg | ND | Jan | 2015 | >8 | >8 | M | Neurology |
| PH-1165-15 | Urine | <i>K. pneumoniae</i> | Pos | Pos | Neg | ND | Jan | 2015 | >8 | >8 | M | Pulmonary Disease |
| PH-1166-15 | Urine | <i>K. pneumoniae</i> | Pos | Pos | Neg | ND | Feb | 2015 | >8 | >8 | M | Surgical ICU |
| PH-1261-15 | Blood | <i>C. freundii</i> | Pos | Pos | Neg | ND | Jul | 2015 | 8 | >8 | M | Nephrology |
| PH-1263-15 | Blood | <i>E. cloacae</i> | Pos | Pos | Neg | ND | Jul | 2015 | >8 | >8 | F | Neonatal ICU |
| PH-1265-15 | Blood | <i>E. cloacae</i> | Pos | Pos | Neg | ND | Jul | 2015 | 8 | 8 | F | Neonatal ICU |
| PH-1266-15 | Blood | <i>E. cloacae</i> | Pos | Pos | Neg | ND | Jul | 2015 | >8 | >8 | F | Neonatal ICU |
| PH-1270-15 | Blood | <i>K. pneumoniae</i> | Pos | Pos | Neg | ND | Jan | 2015 | >8 | >8 | F | Female Medical |
| PH-1271-15 | Wound | <i>R. oxytoca</i> | Pos | Pos | Neg | ND | Jan | 2015 | 0 | >8 | M | Neurology |
| PH-1280-15 | Urine | <i>C. freundii</i> | Pos | Pos | Neg | ND | Jan | 2015 | >8 | >8 | M | Neurology |
| PH-1363-15 | Wound | <i>P. aeruginosa</i> | Pos | Pos | Neg | ND | Sep | 2015 | >8 | >8 | M | Surgical ICU |
| PH-1379-15 | Blood | <i>K. pneumoniae</i> | Pos | Pos | Neg | ND | Sep | 2015 | >8 | >8 | M | Medical ICU |
| PH-1384-15 | Blood | <i>K. pneumoniae</i> | Pos | Pos | Neg | ND | Oct | 2015 | >8 | >8 | F | Medical ICU |
| PH-1394-15 | Wound | <i>K. pneumoniae</i> | Pos | Pos | Neg | ND | Oct | 2015 | >8 | >8 | M | Surgical ICU |
| PH-1419-15 | Urine | <i>K. pneumoniae</i> | Pos | Pos | Neg | ND | Oct | 2015 | >8 | >8 | F | Female Medical |
| PH-1477-15 | Wound | <i>E. cloacae</i> | Pos | Pos | Neg | ND | Oct | 2015 | >8 | >8 | F | Medical ICU |
| PH-1478-15 | Wound | <i>E. cloacae</i> | Pos | Pos | Neg | ND | Oct | 2015 | >8 | >8 | F | Female Medical |
| PH-1482-15 | Wound | <i>K. pneumoniae</i> | Pos | Pos | Neg | ND | Oct | 2015 | >8 | >8 | F | Medical ICU |
| PH-1499-15 | Wound | <i>E. cloacae</i> | Pos | Pos | Neg | ND | Oct | 2015 | >8 | >8 | F | Medical ICU |

Initial empirical therapy

The choice depends on:
Localization and conditions of infection
The presence of risk factors for MDR flora
Severity of the condition
Local Epidemiology

There is an effect:
AB continuation

48-72 hours
Evaluation of the effectiveness
of AB

No effect:
neobhod AB correction

Gain starting
empirical therapy

No results:
empirical therapy

There are results
microbiolog. research

Adding AB
vs MRSA

Addition of AB with
activity against Gr-

Change AB

Targeted therapy

The choice depends on:
Localization and conditions of infection
The presence of risk factors for MDR flora
Severity of the condition
Local Epidemiology

MDR RISK FACTORS OF AGENTS

ESBL extended-spectrum β-lactamase (ESBL)

Prior therapy with cephalosporins or fluoroquinolones

Hospitalization in the previous 3 months.

Being in the ICU

Long hospitalization

CRE Carbapenem-resistant Enterobacteriaceae

Colon colonization CRE

Prior Carbapenema Therapy

CRE high in ICU

Acinetobacter baumannii

Being in the ICU

Prior therapy with wide-spectrum carbapenems and AB

Pseudomonas aeruginosa

Mechanical ventilation of the lungs> 4 days

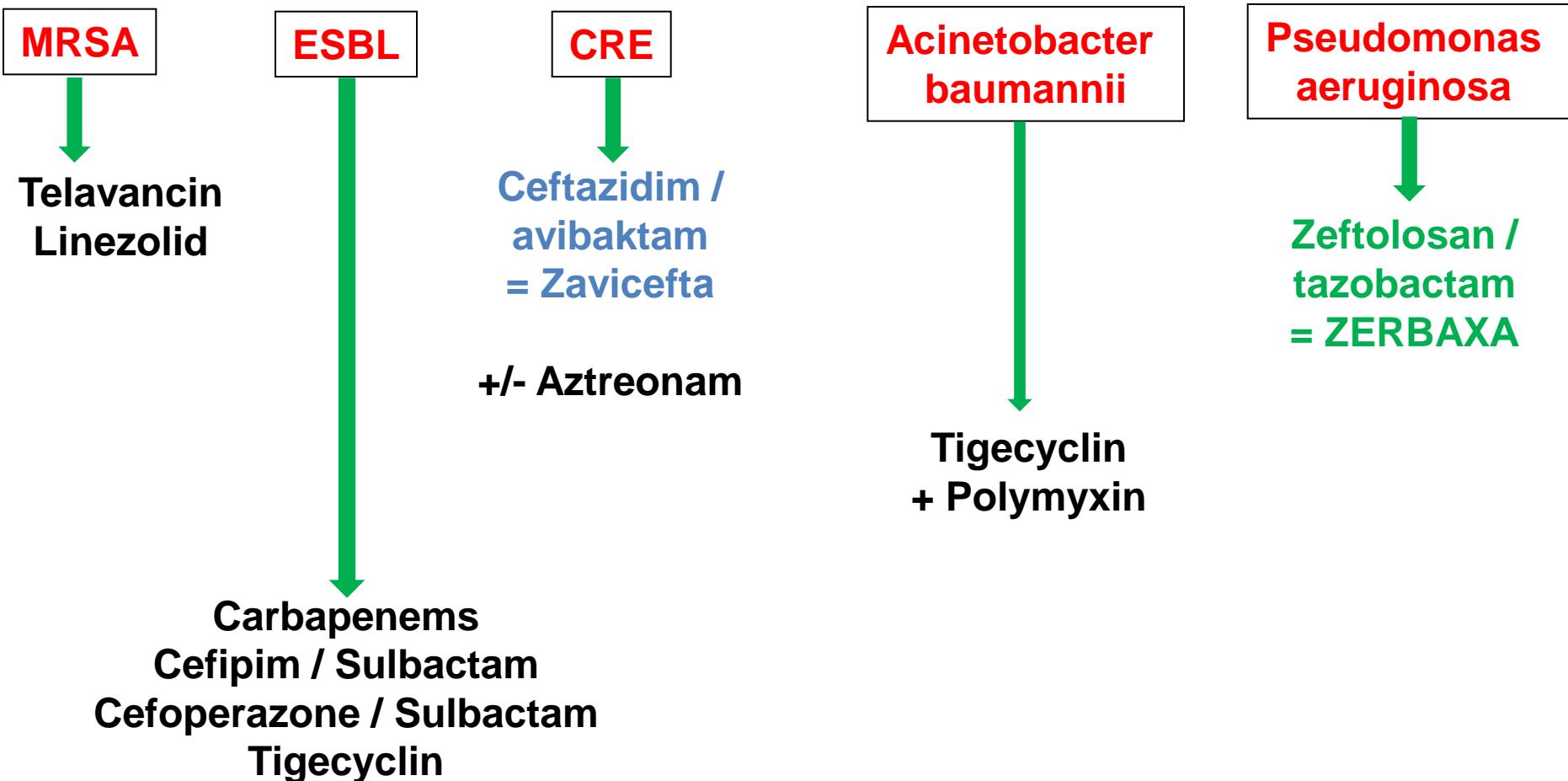
Preceding broad spectrum AB

Glucocorticoidas

Sternotomy

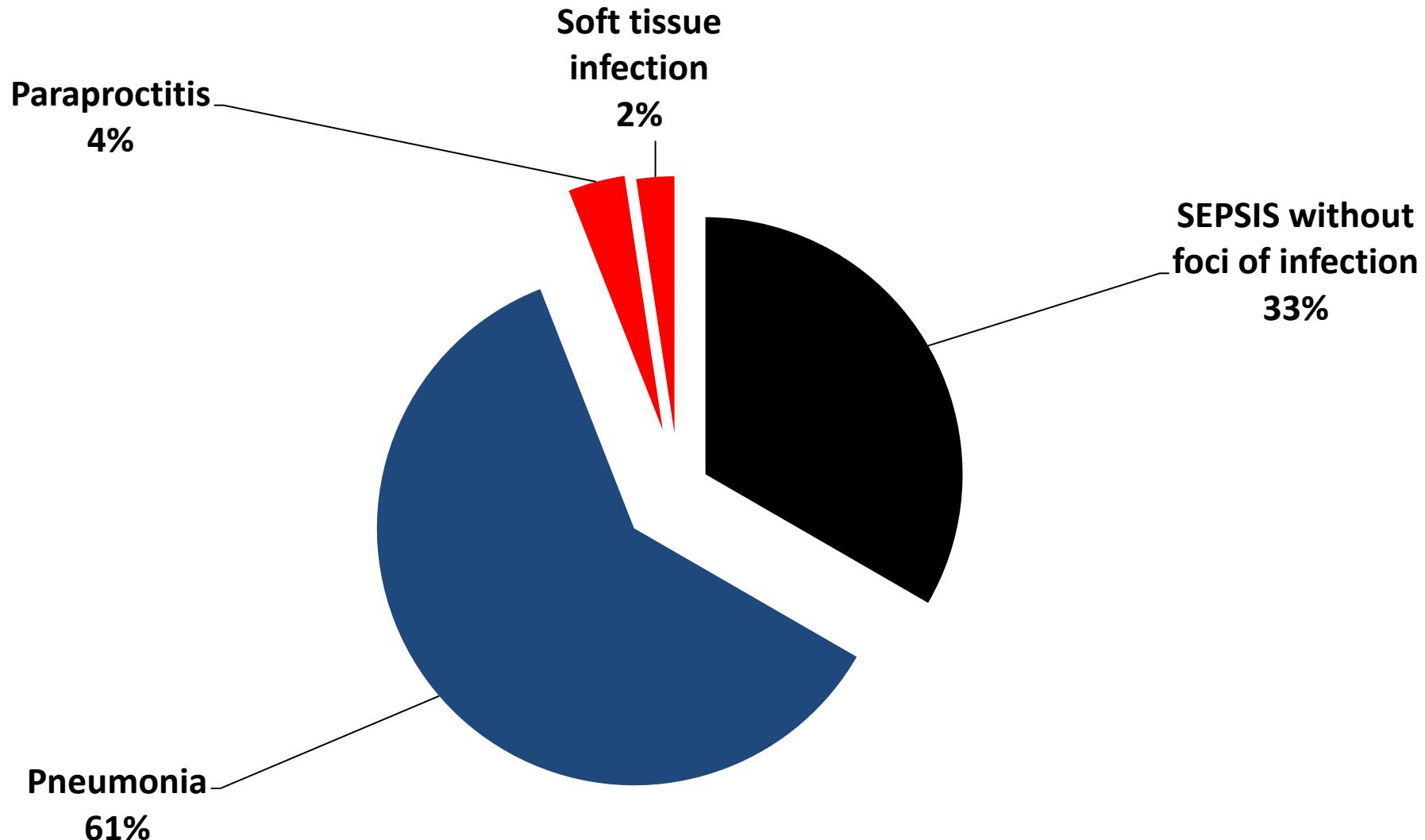
Chronic obstructive pulmonary disease, bronchiectasis, cystic fibrosis

Nosocomial infection - type IIIB – РИСК MRSA, ESBL, CRE, *Acinetobacter baumannii*, *Pseudomonas aeruginosa*



Focus of infection

Infectious complications. Source of infection. 2018 (%)



Soft tissue infection



Antibiotics

Infusion therapy

**Intravenous
immunoglobulins**

Biopsy

Ultrasound

Surgeon observation

Soft tissue infection



Central catheter - Bacteremia *Klebsiella pneumoniae*
Biopsy - *Klebsiella pneumoniae*

Tienam - R

Tigacyclin - ?

Colistin - S

Infusion therapy
130%

Pentaglobin

Artificial nutrition

Biopsy - ?

Ultrasound

Soft tissue infection

Balance correction
KHS correction
Correction of coagulopathy
Hepatoprotectors
Prevention of gastrointestinal paresis
CRRT, Plasmapheresis



Norepinephrine
Dopamine
Hydrocortisone
Cytokinepheresis
LPS-sorption



Soft tissue infection - after medical procedures



Pseudomonas aeruginosa



Soft tissue infection - after medical procedures

Klebsiella pneumoniae



Klebsiella pneumoniae KPC+ Bacteremia + Septicemia + Wound infection



Diagnosis: AML

**D + 30, repeated,
allogeneic, unrelated**

Pancytopenia

***Klebsiella pneumoniae* KPC+ -
chronic carriage in the gastrointestinal tract**

***Klebsiella pneumoniae* KPC+ -
feces, seeding from the oral mucosa**

Septicemia



Septicopyemia screenings in soft tissue - lips

**Central catheter - Bacteremia
Klebsiella pneumoniae KPC+**



Bite wound IV finger of the right hand.

Antibacterial therapy:
Tienam - R
Tigacyclin -?
Colistin - S

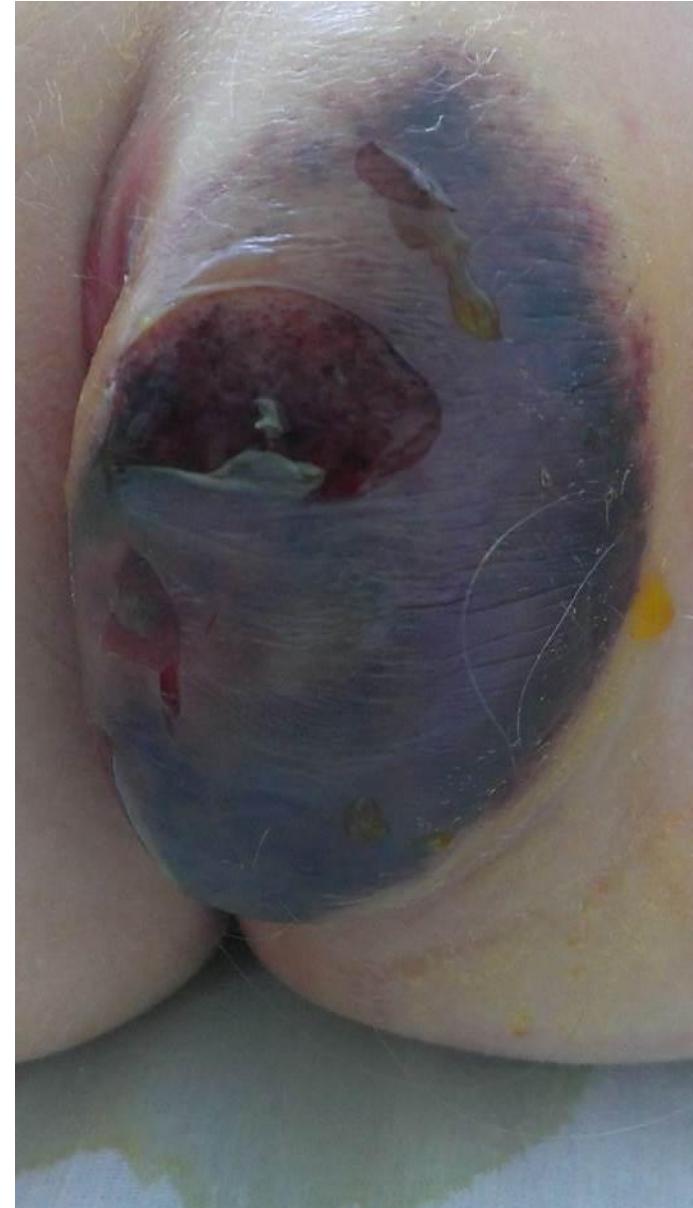
Infection of the soft tissues of the external genital organs



Klebsiella pneumoniae



Infection of the soft tissues of the external genital organs



Pseudomonas aeruginosa

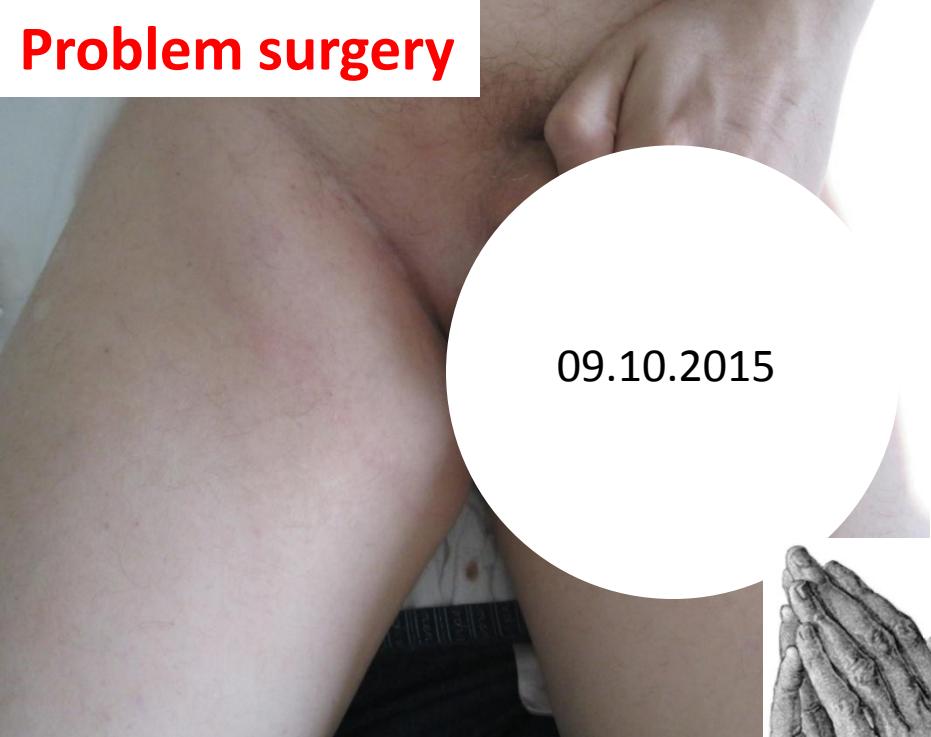
Pseudomonas spp.



**Tienam - R
Colistin - S**

Problem surgery

09.10.2015



11.10.2015





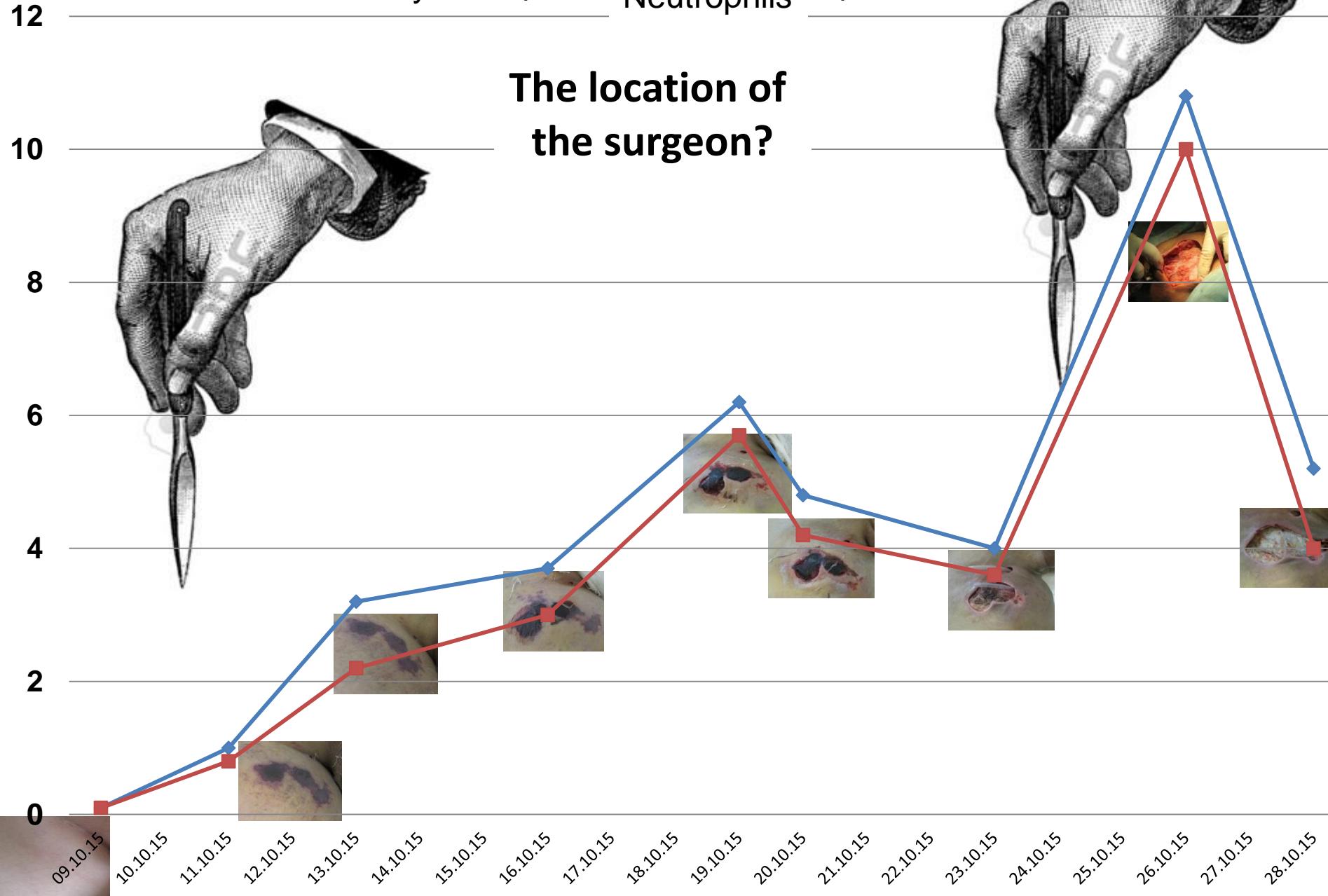




Hematopoiesis Recovery

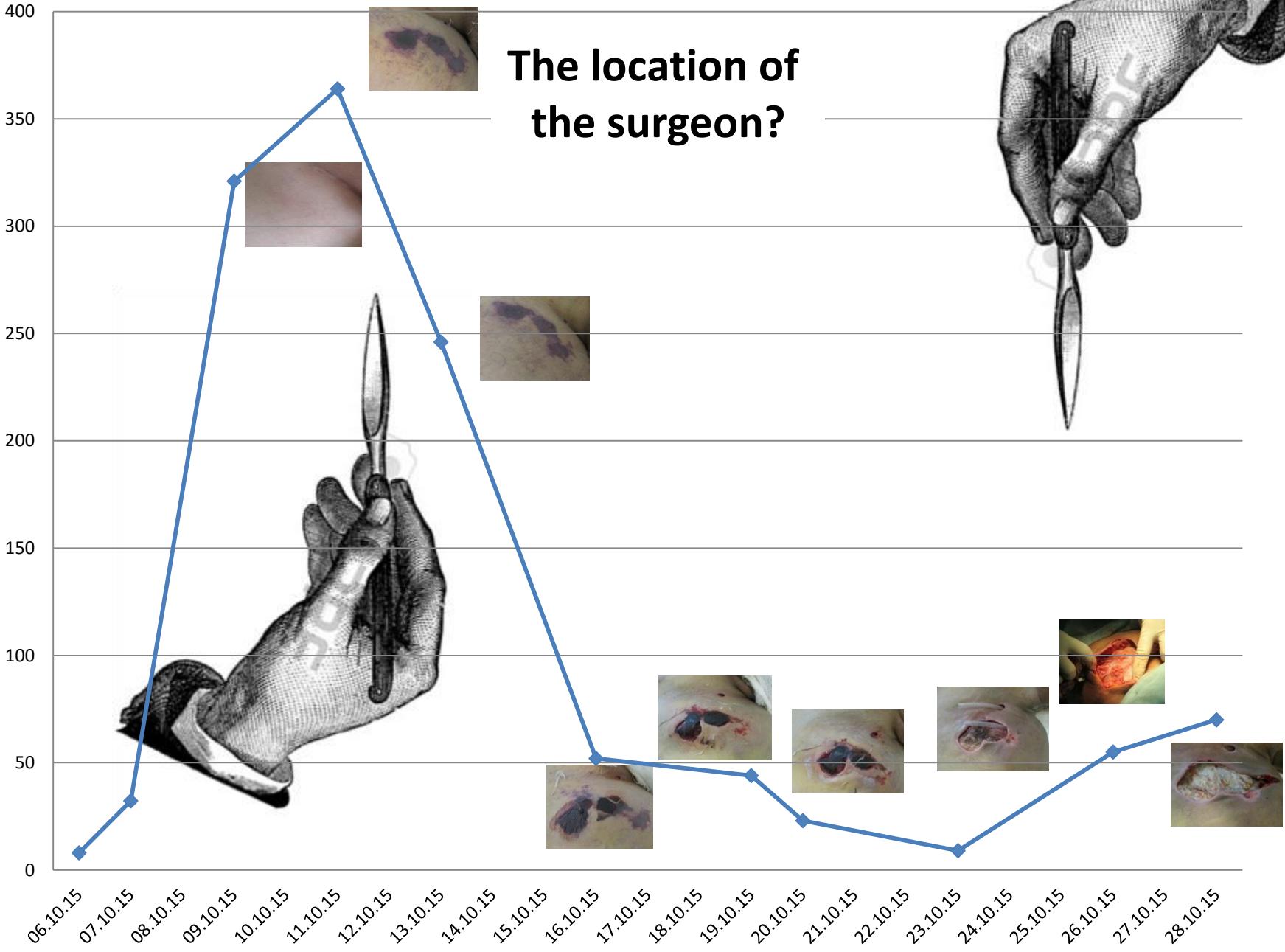
leukocyte VBC) Neutrophils NE) #

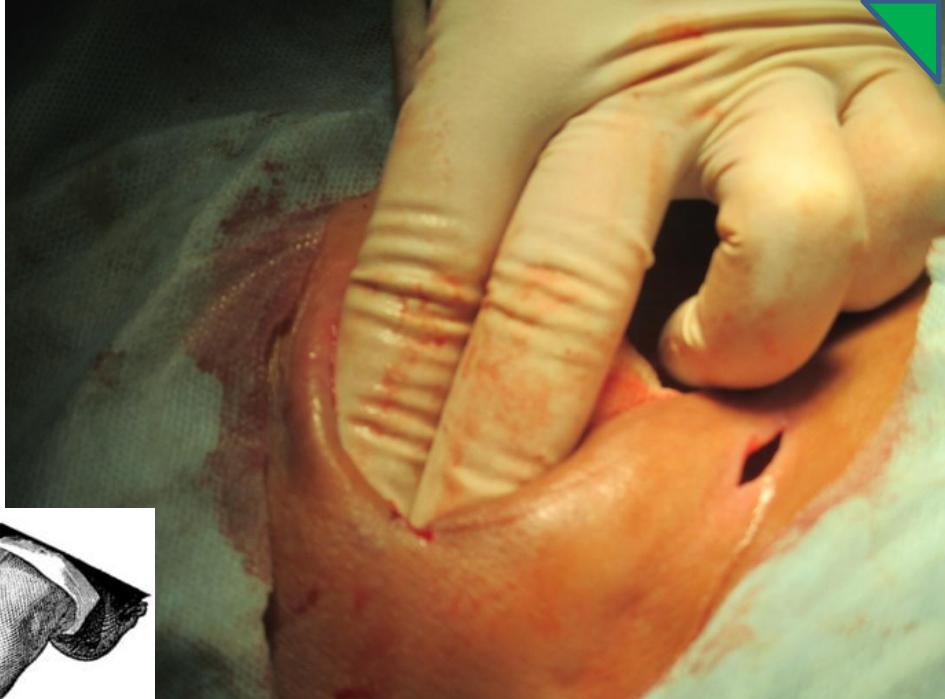
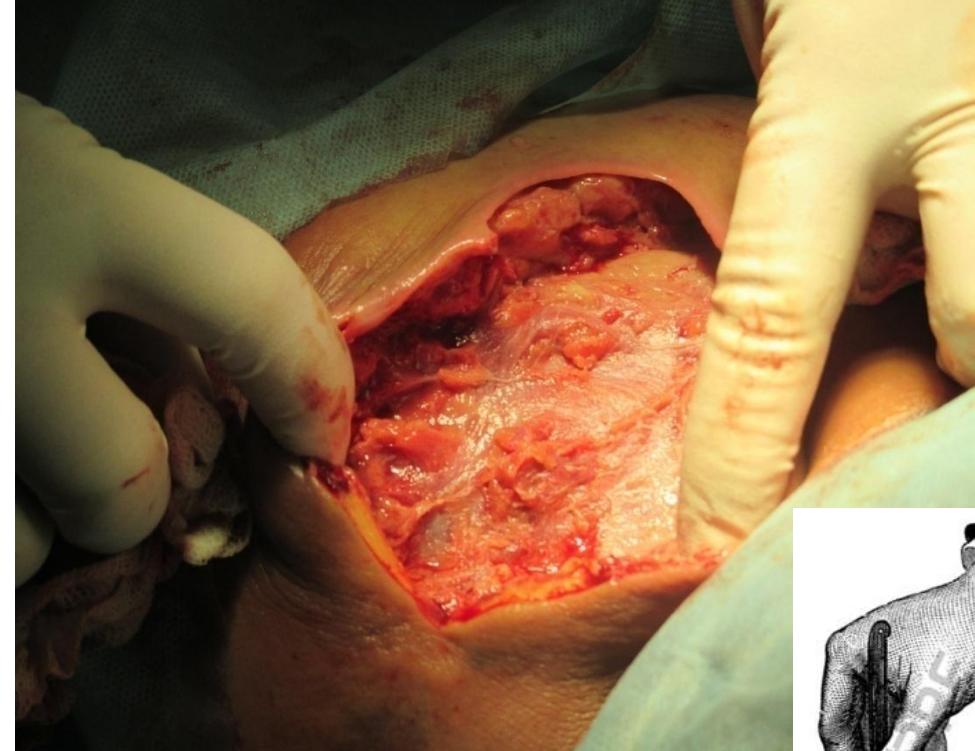
The location of
the surgeon?



CRP

The location of
the surgeon?





26.10.2015





BMT - as a “rescue” therapy

Is soft tissue infection a contraindication to HSCT?

18 year old woman

Diagnosis

Acquired idiopathic extra-severe aplastic anemia

Allogeneic related bone marrow transplantation

Soft tissue infection of the left hand - Pseudomonas aeruginosa

Chronic tonsillitis associated with Pseudomonas aeruginosa.

Chronic rhinosinusitis associated with Pseudomonas aeruginosa.

Therapy 1

Imipenem + Cilastatin 500 mg №4

Fosfomycin 3 gr №4

T>38

CRP>100

PCT>2

Therapy 2

Imipenem + Cilastatin 500 mg №4

Colistin 80 mg №2

Before BMT



$L=0,2 \times 10^9/l$

D+25

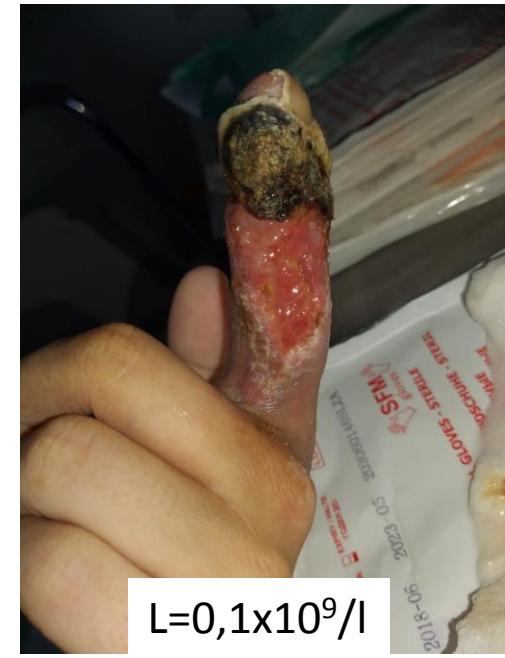
Before BMT



$L=0,1 \times 10^9/l$

D+35

D+15



$L=0,1 \times 10^9/l$

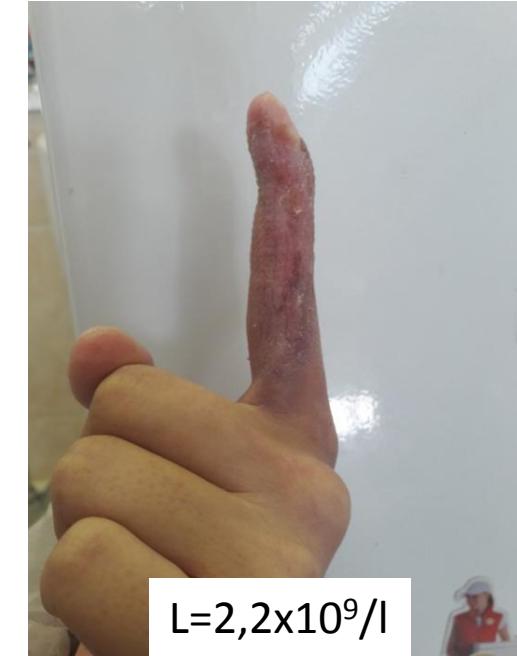
D+60



$L=0,5 \times 10^9/l$



$L=1,1 \times 10^9/l$



$L=2,2 \times 10^9/l$

Hemorrhagic stomatitis

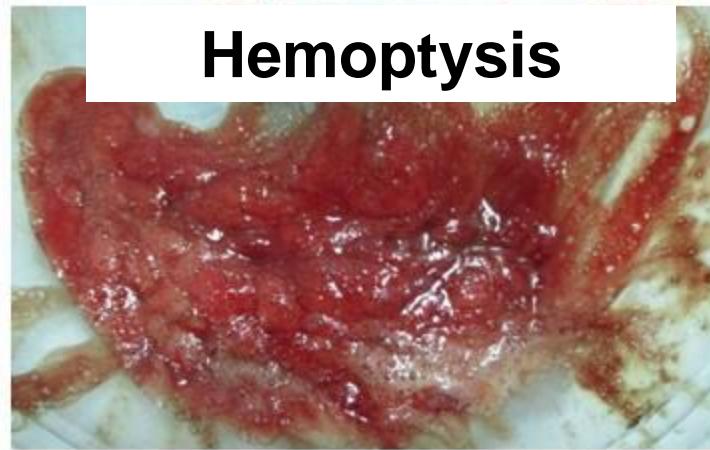


2 days

**Stenotrophomonas
maltophilia**

+ SIRS)

Hemoptysis



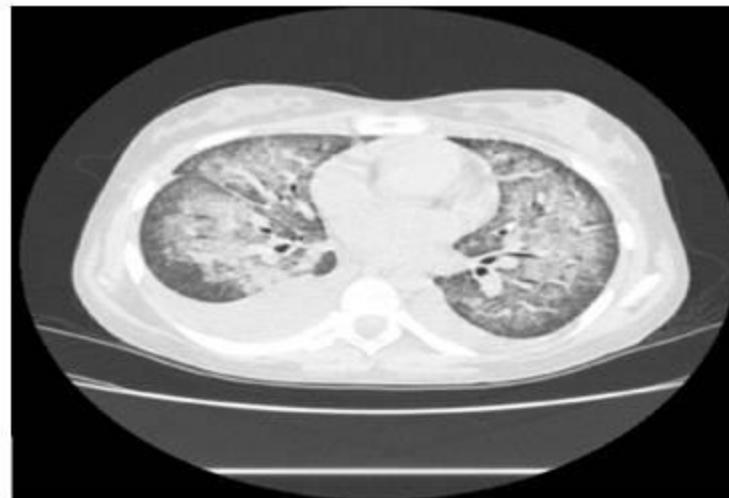
Cough

+

Respiratory insufficiency



1 days



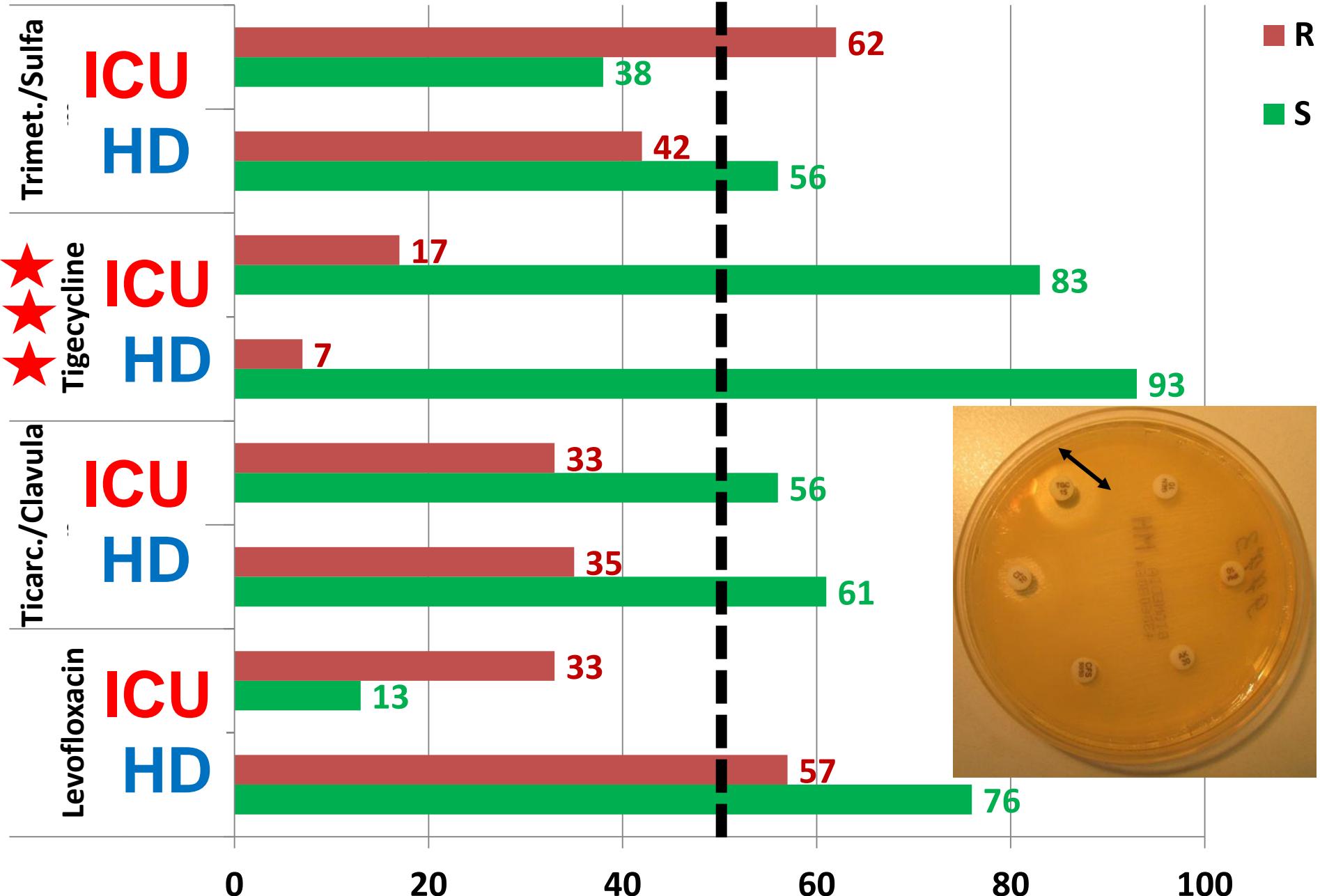
= Stenotrophomonas maltophilia

)

)

125 isolates

Stenotrophomonas maltophilia



EXOTIC? DO NOT THINK! JUST NEED TO BE EXCLUDED

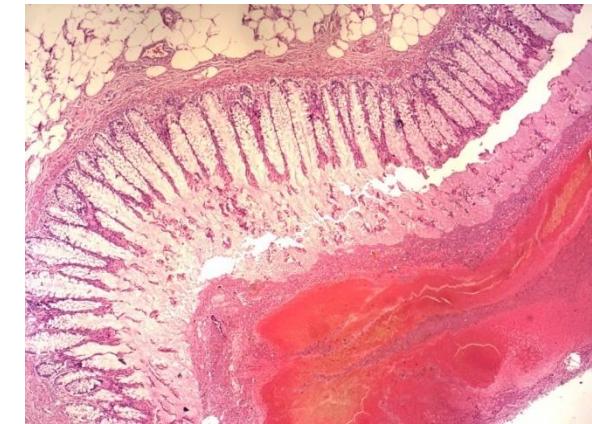
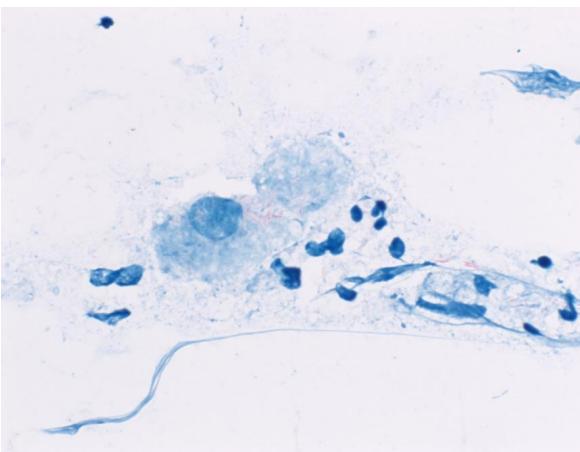
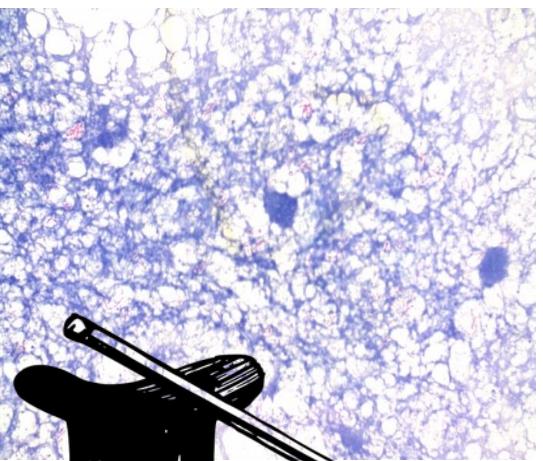
Tuberculosis



Mycobacteriosis

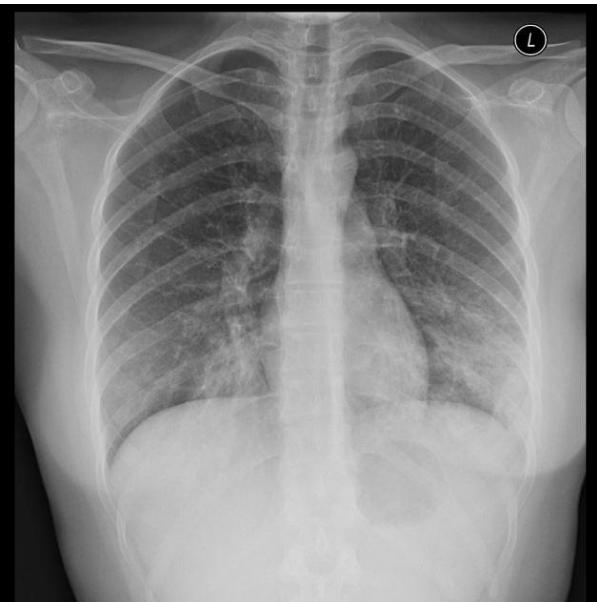


Pseudomembranous colitis

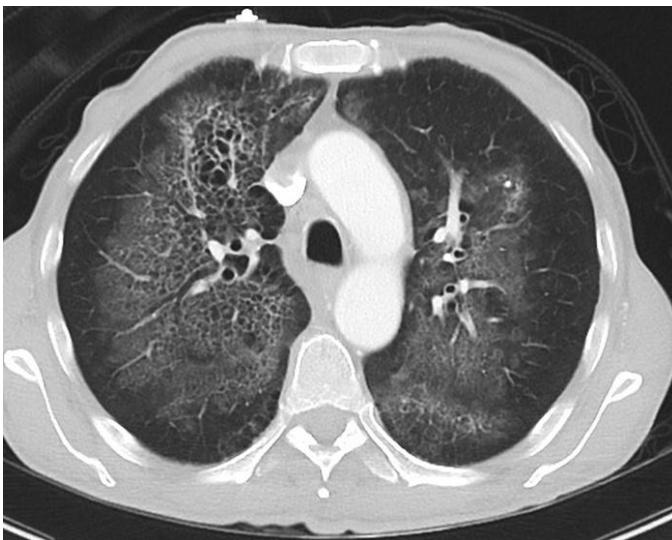


EXOTIC? DO NOT THINK! JUST NEED TO BE EXCLUDED

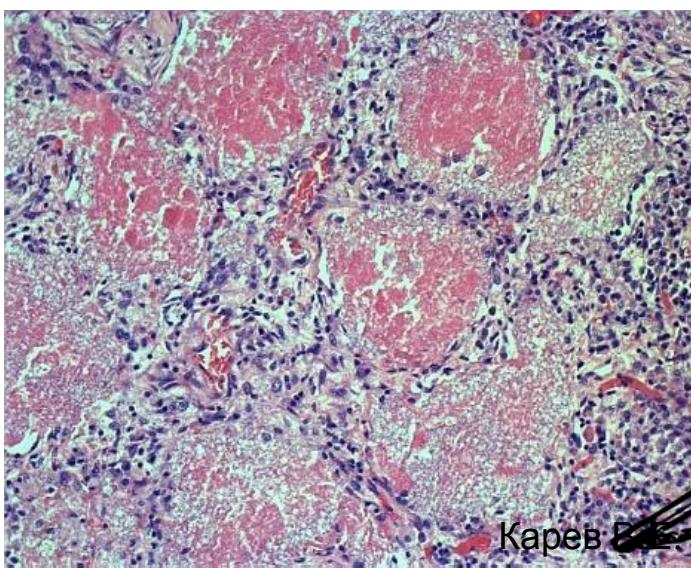
Mycoplasma pneumonia



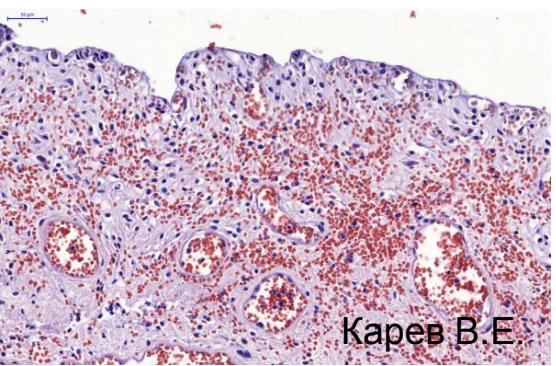
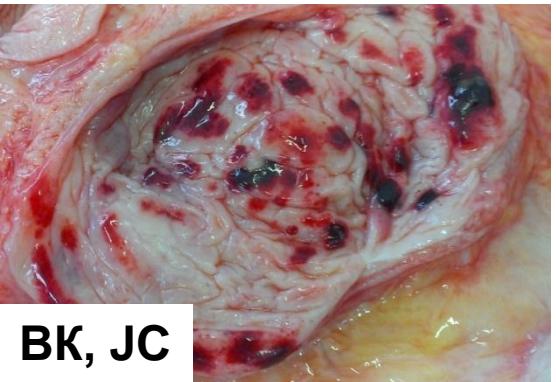
Pneumocystis pneumonia



Chlamydia pneumonia



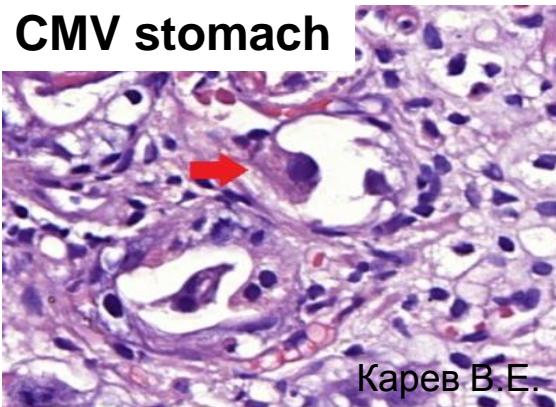
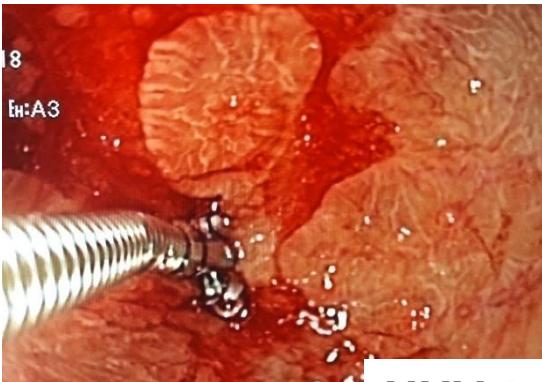
Hemorrhagic cystitis 14-
17%



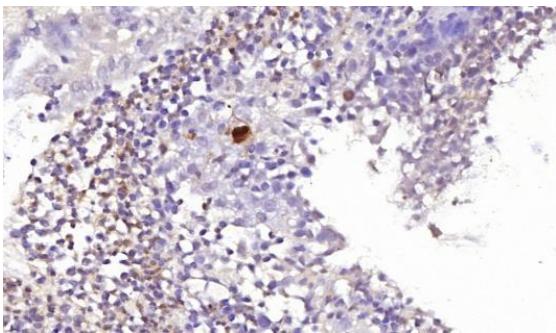
Desquamative
hemorrhagic cystitis
with the formation of ulcers



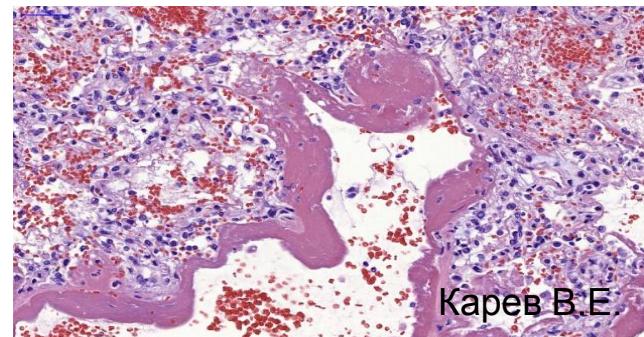
Hemorrhagic enteritis,
colitis



Adeno colon



Diffuse alveolar
bleeding



Interstitial lesions, hyaline
membranes, alterative
changes in the epithelium

VZV





BIOPSY





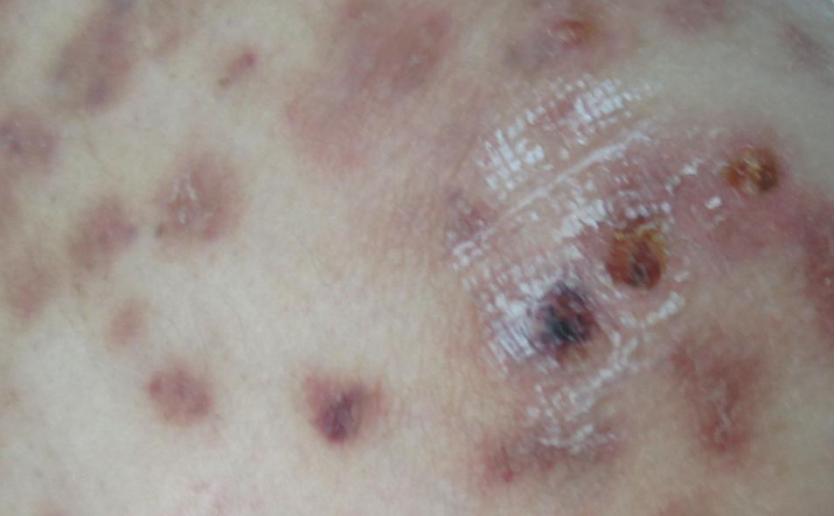
Sepsis

Sepsis markers +

Microbiology - negative

PCR – Vir - negative

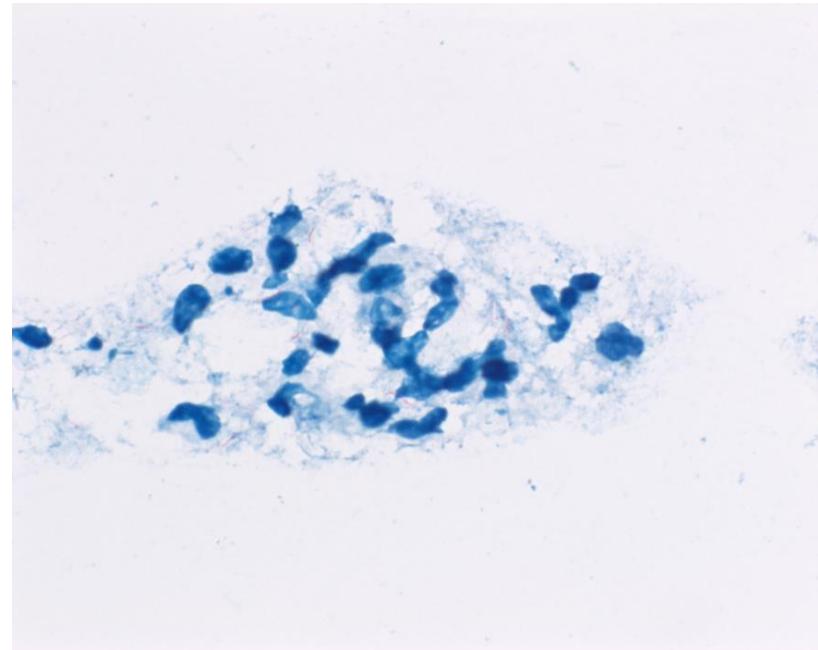
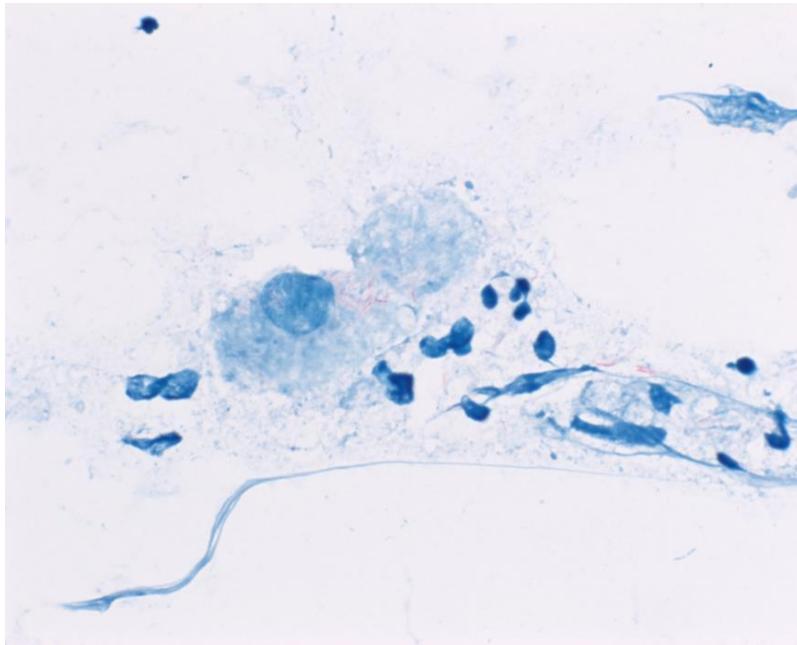
Mycology - negative



There is pus - no pathogen?



Mycobacteriosis



Microscopic method - Coloring according to Zill-Nielsen

Meropenem - S

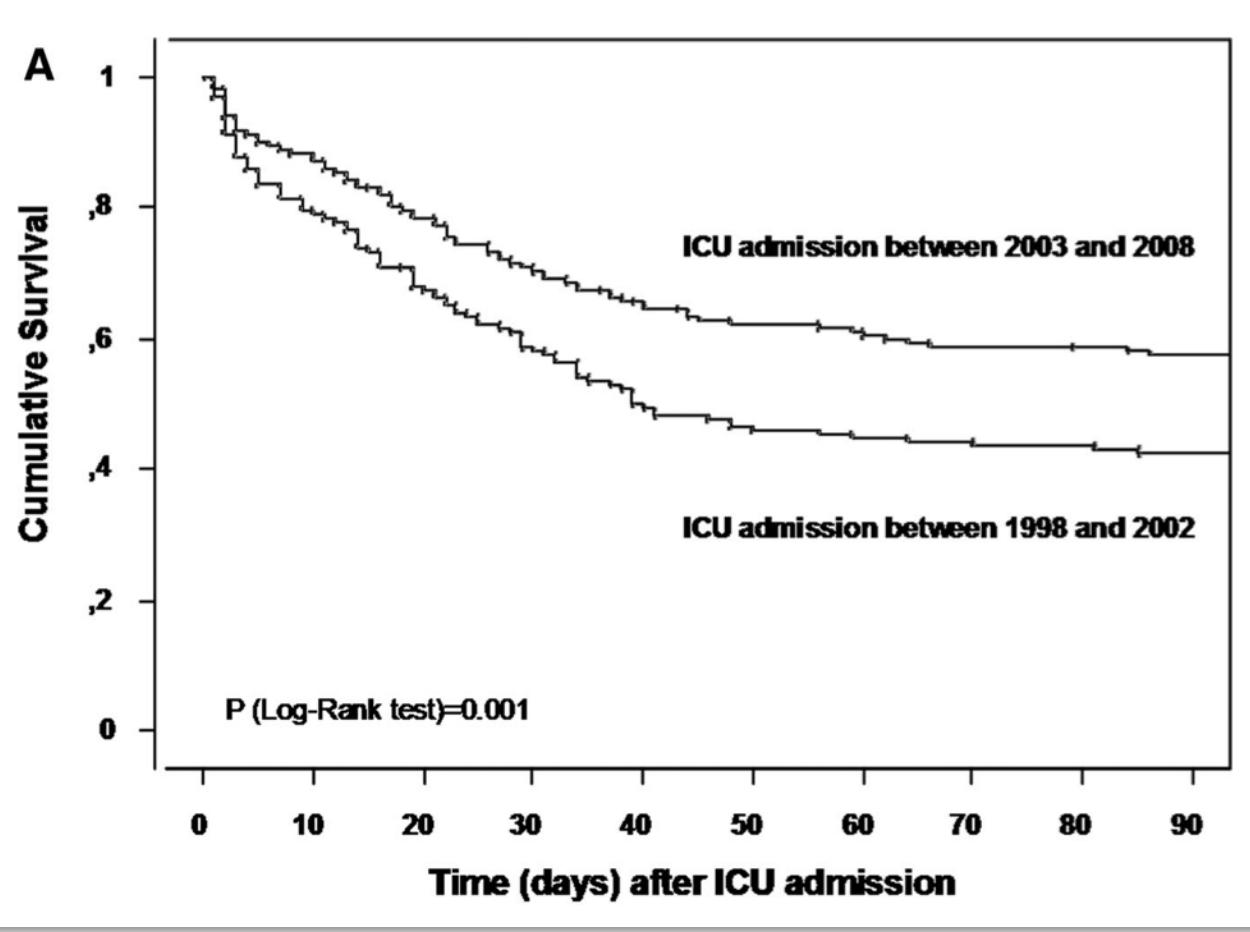
Linezolid - S

Levofloxacin - S

Survival in neutropenic patients with severe sepsis or septic shock

Legrand et al, Crit care med 2012;40:43

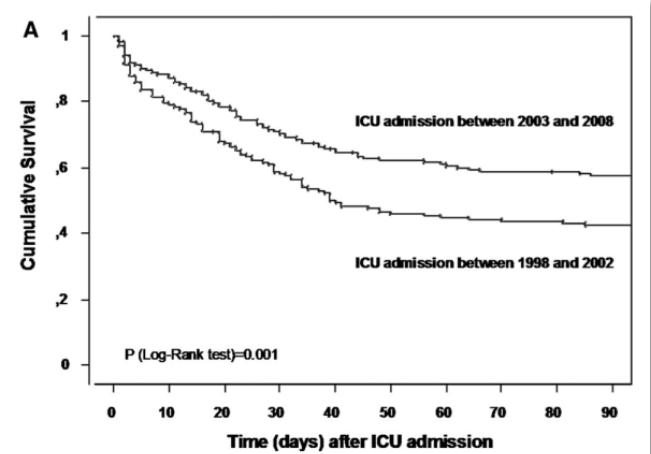
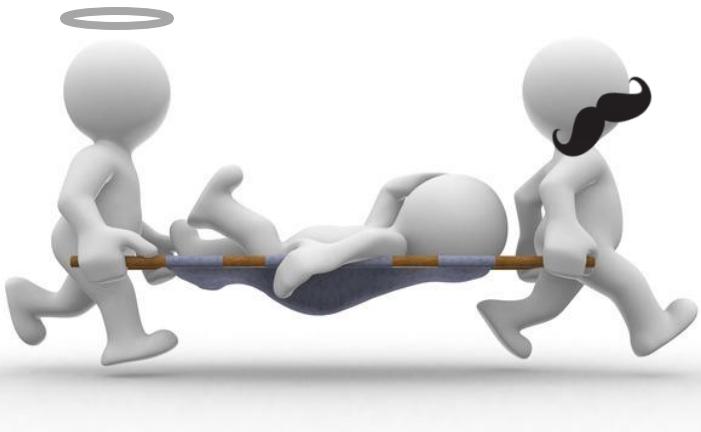
What happened after 2003?



Survival in neutropenic patients with severe sepsis or septic shock

Legrand et al, Crit care med 2012;40:43

Nothing special...



1 - Early translation into intensive care with unproved sepsis

2- Early combined antibacterial therapy in patients with unproved sepsis

3- Early removal of CVV in patients with unproved sepsis

TO CHANGE SEPSIS THERAPY? WHY CHOOSE?

Point last
opportunities!

ICU

Point of non-refund or
do what you want!

ICU

Point FIRST OPPORTUNITY!



The wonderful climate of the Philippines!



Thank you for the attention!